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DB, THE RECEPTOR FOR LEPTIN, NUCLEIC ACIDS
ENCODING THE RECEPTOR, AND USES THEREOF

02/586594



FIELD OF THE INVENTION

The present invention relates to identification of a receptor for a satiety factor, which is involved in body weight homeostasis. Mutations in this receptor are associated with obese phenotypes. In particular, the present invention relates to identification and characterization of the receptor for leptin, including a naturally occurring soluble form of the receptor that is expected to modulate leptin activity, in particular to agonize leptin activity. The invention further relates to the nucleic acids encoding the receptor, and to methods for using the receptor, *e.g.*, to identify leptin analogs, therapeutically, or diagnostically.

BACKGROUND OF THE INVENTION

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Obesity, defined as an excess of body fat relative to lean body mass, is associated with important psychological and medical morbidities, the latter including hypertension, elevated blood lipids, and Type II or non-insulin-dependent diabetes mellitus (NIDDM). There are 6-10 million individuals with NIDDM in the U.S., including 18% of the population of 65 years of age [Harris *et al.*, *Int. J. Obes.*, 11:275-283 (1987)]. Approximately 45% of males and 70% of females with NIDDM are obese, and their diabetes is substantially improved or eliminated by weight reduction [Harris, *Diabetes Care*, 14(3):639-648 (1991)]. As described below, both obesity and NIDDM are strongly heritable.

The assimilation, storage, and utilization of nutrient energy constitute a complex homeostatic system central to survival of metazoa. Among land-dwelling mammals, storage in adipose tissue of large quantities of metabolic fuel as triglycerides is crucial for surviving periods of food deprivation. The need to maintain a fixed level of energy stores without continual alterations in the size and shape of the organism requires the achievement of a balance between energy intake and expenditure.

An individual's level of adiposity is, to a large extent, genetically determined. Examination of the concordance rates of body weight and adiposity amongst mono- and dizygous twins or adoptees and their biological parents have suggested that the heritability of obesity (0.4-0.8) exceeds that of many other traits commonly
 5 thought to have a substantial genetic component, such as schizophrenia, alcoholism, and atherosclerosis [Stunkard *et al.*, *N. Engl. J. Med.*, **322**:1483-1487 (1990)]. Familial similarities in rates of energy expenditure have also been reported [Bogardus *et al.*, *Diabetes*, **35**:1-5 (1986)]. Genetic analysis in geographically delimited populations has suggested that a relatively small number
 10 of genes may account for the 30-50% of variance in body composition [Moll *et al.*, *Am. J. Hum. Genet.*, **49**:1243-1255 (1991)].

Rodent models of obesity include seven apparently single-gene mutations. The most intensively studied mouse obesity mutations are the *ob* (obese) and *db*
 15 (diabetes) genes. When present on the same genetic strain background, *ob* and *db* result in indistinguishable metabolic and behavioral phenotypes, suggesting that these genes may function in the same physiologic pathway [Coleman *et al.*, *Diabetologia*, **14**:141-148 (1978)]. Mice homozygous for either mutation are hyperphagic and hypometabolic, leading to an obese phenotype that is notable at
 20 one month of age. The weight of these animals tends to stabilize at 60-70 g (compared with 30-35 g in control mice). *ob* and *db* animals manifest a myriad of other hormonal and metabolic changes that had made it difficult to identify the primary defect attributable to the mutation [Bray *et al.*, *Am. J. Clin. Nutr.*, **50**:891-902 (1989)]. As noted below, identification of the *OB* gene led to an
 25 understanding of one molecular element.

Each of the rodent obesity models is accompanied by alterations in carbohydrate metabolism resembling those in Type II diabetes in man. In some cases, the severity of the diabetes depends in part on the background mouse strain [Leiter,
 30 *Endocrinology*, **124**:912-922 (1989)]. For both *ob* and *db*, congenic C57BL/Ks mice develop a severe diabetes with ultimate β cell necrosis and islet atrophy,

resulting in a relative insulinopenia. Conversely, congenic C57BL/6J *ob* and *db* mice develop a transient insulin-resistant diabetes that is eventually compensated by β cell hypertrophy resembling human Type II diabetes.

- 5 The phenotype of *ob* and *db* mice resembles human obesity in ways other than the development of diabetes - the mutant mice eat more and expend less energy than do lean controls (as do obese humans). This phenotype is also quite similar to that seen in animals with lesions of the ventromedial hypothalamus, which suggests that both mutations may interfere with the ability to properly integrate or respond
- 10 to nutritional information within the central nervous system. Support for this hypothesis comes from the results of parabiosis experiments [Coleman, *Diabetologia*, 9:294-298 (1973)] that suggest *ob* mice are deficient in a circulating satiety factor and that *db* mice are resistant to the effects of the *ob* factor (possibly due to an *ob* receptor defect). These experiments have led to the conclusion that
- 15 obesity in these mutant mice may result from different defects in an afferent loop and/or integrative center of the postulated feedback mechanism that controls body composition.

Using molecular and classical genetic markers, the *ob* and *db* genes have been

20 mapped to proximal chromosome 6 and midchromosome 4, respectively [Bahary *et al.*, *Proc. Nat. Acad. Sci. USA*, 87:8642-8646 (1990); Friedman *et al.*, *Genomics*, 11:1054-1062 (1991)]. In both cases, the mutations map to regions of the mouse genome that are syntenic with human, suggesting that, if there are human homologs of *ob* and *db*, they are likely to map, respectively, to human

25 chromosomes 7q and 1p. Defects in the *db* gene may result in obesity in other mammalian species: in genetic crosses between Zucker *fa/fa* rats and Brown Norway *+/+* rats, the *fa* mutation (rat chromosome 5) is flanked by the same loci that flank *db* in mouse [Truett *et al.*, *Proc. Natl. Acad. Sci. USA*, 88:7806-7809 (1991)].

A major advance in understanding the molecular basis for obesity occurred with the cloning of the *ob* gene. The mouse obesity (*ob*) gene encodes an adipose tissue-derived signaling factor for body weight homeostasis [Zhang *et al.*, *Nature*, 372:425 (1994); U.S. Patent Application No. 08/292,345 filed August 17, 1994; 5 U.S. Patent Application No. 08/483,211, filed June 7, 1995, each of which is hereby incorporated by reference in its entirety]. Several recent studies have shown that recombinant OB protein (leptin) purified from *Escherichia coli* can correct the obesity related phenotypes in *ob/ob* mice when exogenously administered [Campfield *et al.*, *Science*, 269:546 (1995); Pellymounter *et al.*, 10 *Science*, 269:540, (1995); Halaas *et al.*, *Science*, 269:543 (1995); Stephens *et al.*, *Nature*, 377:530 (1995)]. Weight-reducing effects of recombinant leptin were also observed in normal mice and mice with diet-induced obesity. Although the target tissues that mediate the effects of leptin have not yet been defined, the instant inventors have predicted the brain as a target of leptin activity. Indeed, the work 15 of Campfield *et al.* (*supra*) and Stephens *et al.* (*supra*) demonstrates that leptin introduced into the lateral or third brain ventricle is effective at low doses, arguing for a direct central effect of the leptin molecule.

Recent studies have suggested that obese humans and rodents (other than *ob/ob* 20 mice) are not defective in their ability to produce leptin mRNA or protein and generally produce higher levels than lean individuals [Maffei *et al.*, *Nature Med.*, 1:1155 (1995); Considine *et al.*, *J. Clin. Invest.*, 95:2986 (1995); Lonnqvist *et al.*, *Nature Med.*, 1:950 (1995); Hamilton *et al.*, *Nature Med.*, 1:953 (1995)]. These data suggest that resistance to normal or elevated levels of leptin may be important 25 factors in human obesity. However, a recent report of identification of a leptin receptor did not identify any mutations in the *ob* allele [Tartaglia *et al.*, *Cell*, 83:1263-1271 (1995)].

Accordingly, there is a need in the art to identify a receptor for leptin.

There is a further need to characterize mutations in the leptin receptor, particularly as they may be associated with obesity.

There is a still further need to identify and characterize functions of the leptin receptor, or variants thereof.

These and other needs in the art are addressed by the present invention.

The citation of any reference herein should not be construed as an admission that such a reference is available as prior art to the application.

SUMMARY OF THE INVENTION

The present invention is directed to a leptin receptor (OB-R) polypeptide, nucleic acids encoding such polypeptide, non-coding nucleic acids flanking the coding sequences of the gene, oligonucleotides that hybridize to such nucleic acids, antibodies to the polypeptide, and diagnostic, therapeutic, and cosmetic compositions and methods utilizing the polypeptide, nucleic acids, or antibodies, or combinations thereof.

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Thus, in a first aspect of the invention, the leptin receptor (also termed herein OB receptor or OB-R) is characterized by specific binding to leptin under physiological conditions; expression at high levels in cells of the hypothalamus, and expression at lower levels in adipose tissue, testes, heart, and brain; and having sequence similarity to gp130 cytokine receptors. In another embodiment, the leptin receptor is encoded by a nucleic acid which is identifiable with a polymerase chain reaction (PCR) probe selected from group consisting of a probe for clone 7 (forward primer SEQ ID NO:42 and reverse primer SEQ ID NO:43), a probe for clone 11 (forward primer SEQ ID NO:44 and reverse primer SEQ ID NO:45), and both clone 7 and clone 11. In a specific embodiments, leptin receptor is encoded by a nucleic acid which is identifiable with a PCR probe

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selected from the group consisting of a probe for clone 42 (forward primer SEQ ID NO:26 and reverse primer SEQ ID NO:46); a probe for clone 46 (forward primer SEQ ID NO:47 and reverse primer SEQ ID NO:48); a probe for clone 58 (forward primer SEQ ID NO:49 and reverse primer SEQ ID NO:50); a probe for clone S14 (forward primer SEQ ID NO:51 and reverse primer SEQ ID NO:52); and a probe for clone S3 (forward primer SEQ ID NO:53 and reverse primer SEQ ID NO:54).

In specific Examples, *infra*, the leptin receptor is selected from the group consisting of OB-Ra (SEQ ID NO:2), OB-Rb (SEQ ID NO:4), OB-Rc (SEQ ID NO:6), OB-Rd (SEQ ID NO:8), and OB-Re (SEQ ID NO:10), or allelic variants thereof. Alternatively, the leptin receptor may have a sequence selected from the group consisting of:

N-terminal corresponding to OB-Ra through Lys⁸⁸⁹ and C-terminal corresponding to a C-terminal selected from the group consisting of OB-Rb, OB-Rc, and OB-Rd after Lys⁸⁸⁹;

N-terminal corresponding to OB-Rb or OB-Rc through Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra or OB-Rd after Lys⁸⁸⁹;

N-terminal corresponding to OB-Rd through Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra, OB-Rb, or OB-Rc;

N-terminal corresponding to OB-R from Pro⁶⁶⁴ to Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra, OB-Rb, OB-Rc, and OB-Rd; and

N-terminal corresponding to OB-R from Met⁷³³ to Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra, OB-Rb, OB-Rc, and OB-Rd; and

N-terminal selected from the group consisting of OB-Ra, OB-Rb, OB-Rd, and OB-R from Pro⁶⁶⁴ to His⁷⁹⁶, and OB-Re from His⁷⁹⁶;

N-terminal selected from the group consisting of OB-Ra, OB-Rb, OB-Rd, and OB-R from Met⁷³³ to His⁷⁹⁶, and OB-Re from His⁷⁹⁶, or allelic variants thereof.

In another embodiment, leptin receptor may have an N-terminal sequence is selected from the group consisting of

- amino acid residues 1-889;
- amino acid residues 23-889;
- 5 amino acid residues 28-889;
- amino acid residues 133-889;
- amino acid residues 733-889;
- amino acid residues 1-796;
- amino acid residues 23-796;
- 10 amino acid residues 28-796;
- amino acid residues 133-796; and
- amino acid residues 733-796; and

and a C-terminal sequence is selected from the group consisting of SEQ ID NO:11;

- 15 SEQ ID NO:12; SEQ ID NO:13; SEQ ID NO:14; and SEQ ID NO:15, wherein the numbering is based on the amino acid sequence of the full length transcribed murine leptin receptor, including the signal peptide, or allelic variants thereof.

- In a specific embodiment, the leptin receptor is a soluble receptor. Such a soluble
- 20 receptor may be selected from the group consisting of OB-Re; an N-terminal sequence which selected from the group consisting of OB-Ra, OB-Rb, OB-Rd, and OB-R from Pro⁶⁶⁴ to His⁷⁹⁶, and a C-terminal sequence which is OB-Re from His⁷⁹⁶; and OB-R from Met⁷³³ to His⁷⁹⁶, and a C-terminal sequence which is OB-Re from His⁷⁹⁶; an N-terminal sequence which is selected from the group consisting of
 - 25

- amino acid residues 1-796;
- amino acid residues 23-796;
- amino acid residues 28-796;
- amino acid residues 133-796; and
- 30 amino acid residues 733-796; and

a C-terminal sequence which is SEQ ID NO:15; wherein the numbering is based on the amino acid sequence of the full length transcribed murine leptin receptor, including the signal peptide, or allelic variants thereof.

5 Alternatively, the leptin receptor comprises a transmembrane domain, and is an integral membrane protein. In this embodiment, the leptin receptor may further comprise a JAK binding motif selected from "Box 1," "Box 2," and "Box 1" and "Box 2", which motif is downstream of the transmembrane domain.

10 In one specific embodiment, the leptin receptor is a human leptin receptor. In another specific embodiment, exemplified *infra*, the leptin receptor is a murine leptin receptor. In a further specific embodiment, the leptin receptor is a human leptin receptor comprising a divergent amino acid substitution from the corresponding position of the murine leptin receptor. In another embodiment, the
15 leptin receptor is a human leptin receptor comprising conservative amino acid substitutions. In a specific embodiment, conservative amino acid substitutions from murine leptin receptor are made in human leptin receptor. In yet another embodiment, conservative amino acid substitutions that enhance secondary structure, *e.g.*, α -helical propensity, are made.

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The present invention further provides an antigenic fragment of the leptin receptor. In a specific embodiment, the antigenic fragment is selected from the group consisting of SEQ ID NO:32, SEQ ID NO:33, and SEQ ID NO:34.

25 The invention further relates to a derivative of the soluble form of the leptin receptor attached to a chemical moiety. Preferably, the chemical moiety is a water-soluble polymer. More preferably, the water soluble polymer is polyethylene glycol.

30 In another aspect, the invention provides an isolated nucleic acid encoding the leptin receptor, particularly as set forth above. In specific examples, *infra*, the

invention provides cDNA encoding various splice forms of murine leptin receptor. In particular, the present invention provides nucleic acids having sequences corresponding or complementary to SEQ ID NO:1, 3, 5, 7, or 9:

5 More particularly, the invention provides an isolated DNA molecule encoding on expression a leptin receptor polypeptide selected from the group consisting of:

a polypeptide coding sequence of a DNA molecule of SEQ ID NO:1, 3, 5, 7, or 9;

a DNA molecule complementary to the DNA molecule defined in

10 (a);

a DNA molecule which hybridizes to the DNA molecule of (a) or (b), or a hybridizable fragment thereof;

a DNA molecule which is identifiable with a polymerase chain reaction (PCR) probe selected from group consisting of a probe for clone 7 (forward primer SEQ ID NO:42 and reverse primer SEQ ID NO:43), a

15 probe for clone 11 (forward primer SEQ ID NO:44 and reverse primer SEQ ID NO:45), and both clone 7 and clone 11; and

a DNA molecule that codes on expression for the polypeptide encoded by any of the foregoing DNA molecules.

20 Preferably the DNA molecule is human; in specific Examples, *infra*, the DNA molecule is murine. In specific embodiments, the DNA molecule codes on expression for a polypeptide selected from the group consisting of a leptin receptor selected from the group consisting of OB-Ra, OB-Rb, OB-Rc, OB-Rd, and OB-Re, or allelic variants thereof; a leptin receptor selected from the group consisting of:

25 N-terminal corresponding to OB-Ra through Lys⁸⁸⁹ and C-terminal corresponding to a C-terminal selected from the group consisting of OB-Rb, OB-Rc, and OB-Rd after Lys⁸⁸⁹;

N-terminal corresponding to OB-Rb or OB-Rc through Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra or OB-Rd after Lys⁸⁸⁹;

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N-terminal corresponding to OB-Rd through Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra, OB-Rb, or OB-Rc;

N-terminal corresponding to OB-R from Pro⁶⁶⁴ to Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra, OB-Rb, OB-Rc, and OB-Rd;
5 and

N-terminal corresponding to OB-R from Met⁷³³ to Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra, OB-Rb, OB-Rc, and OB-Rd; and

N-terminal selected from the group consisting of OB-Ra, OB-Rb, OB-Rd, and OB-R from Pro⁶⁶⁴ to His⁷⁹⁶, and OB-Re from His⁷⁹⁶;

10 N-terminal selected from the group consisting of OB-Ra, OB-Rb, OB-Rd, and OB-R from Met⁷³³ to His⁷⁹⁶, and OB-Re from His⁷⁹⁶, or allelic variants thereof;

a leptin receptor wherein the N-terminal sequence is selected from the group consisting of

15 amino acid residues 1-889;
amino acid residues 23-889;
amino acid residues 28-889;
amino acid residues 133-889;
amino acid residues 733-889;
20 amino acid residues 1-796;
amino acid residues 23-796;
amino acid residues 28-796;
amino acid residues 133-796; and
amino acid residues 733-796;

25 and the C-terminal sequence is selected from the group consisting of SEQ ID NO:11; SEQ ID NO:12; SEQ ID NO:13; SEQ ID NO:14; and SEQ ID NO:15, wherein the numbering is based on the amino acid sequence of the full length transcribed murine leptin receptor, including the signal peptide, or allelic variants thereof.

The invention further contemplates, as a corollary to the coding nucleic acids described above, an oligonucleotide hybridizable under stringent conditions to the nucleic acid molecule encoding leptin receptor. In specific embodiments, exemplified *infra*, the oligonucleotide is selected from the group consisting of SEQ

5 ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, SEQ ID NO:24, SEQ ID NO:25, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:28, SEQ ID NO:29, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO:35, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID

10 NO:46, SEQ ID NO:47, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, and SEQ ID NO:54. The oligonucleotide may be labeled.

In addition to the coding DNA, the present invention provides vectors comprising

15 such DNA. A vector of the invention may be a cloning vector, or it may be an expression vector which comprises the DNA encoding leptin receptor operatively associated with an expression control sequence. Naturally, the invention extends to an unicellular host transformed or transfected with a DNA molecule, cloning vector, or expression vector of the invention. Such a unicellular host may be

20 selected from the group consisting of bacteria, yeast, mammalian cells, plant cells, and insect cells, in tissue culture. In specific embodiments, the host may be selected from the group consisting of *E. coli*, *Pseudomonas*, *Bacillus*, *Streptomyces*, *Saccharomyces*, *Pichia*, *Candida*, *Hansenula*, *Torulopsis*, CHO, R1.1, B-W, LM, COS 1, COS 7, BSC1, BSC40, BMT10, and Sf9 cells.

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The invention further relates to a recombinant method for preparing a leptin receptor polypeptide comprising culturing a host cell comprising an expression vector of the invention under conditions that provide for expression of the leptin receptor polypeptide; and recovering the expressed polypeptide.

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The invention further provides an antisense nucleic acid that hybridizes with an mRNA encoding leptin receptor, and a ribozyme which cleaves an mRNA encoding a leptin receptor.

- 5 In another embodiment, the invention provides a transgenic vector comprising a DNA molecule encoding leptin receptor, or an expression vector of the invention.

In another aspect, the invention provides an antibody specific for a leptin receptor. The antibody may be a monoclonal or polyclonal antibody. In a specific
10 embodiment, the antibody may be labeled with a detectable label. Naturally, the invention extends to an immortal cell line that produces a monoclonal antibody.

In a specific embodiment, the invention provides a method for preparing an antibody specific for a leptin receptor, comprising: immunizing a host animal
15 with the leptin receptor admixed with an adjuvant; and obtaining antibody from the immunized host animal. In another specific embodiment, exemplified *infra*, the method for preparing an antibody specific for a leptin receptor comprises conjugating a peptide having a sequence selected from the group consisting of SEQ ID NO:32, SEQ ID NO:33, and SEQ ID NO:34 to a carrier protein; immunizing
20 a host animal with the peptide-carrier protein conjugate of step (a) admixed with an adjuvant; and obtaining antibody from the immunized host animal.

In conjunction with the antibodies of the invention, the invention provides a method for measuring the presence of a leptin receptor in a sample, comprising
25 contacting a sample suspected of containing a leptin receptor with an antibody that specifically binds to the leptin receptor under conditions which allow for the formation of reaction complexes comprising the antibody and the leptin receptor; and detecting the formation of reaction complexes comprising the antibody and leptin receptor in the sample, wherein detection of the formation of reaction
30 complexes indicates the presence of leptin receptor in the sample. In a specific embodiment, the antibody is bound to a solid phase support. As a corollary to the

method of measuring the presence of leptin receptor in a sample, the invention provides an *in vitro* method for evaluating the level of leptin receptor in a biological sample comprising detecting the formation of reaction complexes in a biological sample as described; and evaluating the amount of reaction complexes formed, which amount of reaction complexes corresponds to the level of leptin receptor in the biological sample. The invention further relates to an *in vitro* method for detecting or diagnosing the presence of a disease associated with elevated or decreased levels of leptin receptor in a subject comprising evaluating the level of leptin receptor in a biological sample from a subject as described; and comparing the level detected in step (a) to a level of leptin receptor present in normal subjects or in the subject at an earlier time, wherein an increase in the level of leptin receptor as compared to normal levels indicates a disease associated with elevated levels of leptin receptor, and decreased level of leptin receptor as compared to normal levels indicates a disease associated with decreased levels of leptin receptor.

The present invention also provides a pharmaceutical composition comprising a soluble leptin receptor, and a pharmaceutically acceptable carrier. Alternatively, a pharmaceutical composition of the invention may comprise a transgenic vector, *e.g.*, a viral vector or naked DNA, for administration to a subject for gene therapy. Preferably, such a vector is targeted to the brain, more preferably the hypothalamus. The invention further provides a method for treating obesity in a subject comprising administering a therapeutically effective amount of the pharmaceutical composition of the invention. The method of treatment may further comprise administering a treatment for diabetes, high blood pressure, and high cholesterol.

In another embodiment, the invention provides a body appearance improving cosmetic composition for reducing the body weight of an individual comprising a soluble leptin receptor, and an acceptable carrier. The invention further provides

a method for improving the body appearance of an individual comprising administering the cosmetic composition of the invention.

- 5 Accordingly, it is a principal object of the present invention to provide modulators of body weight as defined herein in purified form, that exhibit certain characteristics and activities associated with control and variation of adiposity and fat content of mammals.
- 10 It is a further object of the present invention to provide methods for the detection and measurement of the modulators of weight control as set forth herein, as a means of the effective diagnosis and monitoring of pathological conditions wherein the variation in level of such modulators is or may be a characterizing feature.
- 15 It is a still further object of the present invention to provide a method and associated assay system for the screening of substances, such as drugs, agents and the like, that are potentially effective to either mimic or inhibit the activity of leptin binding to its receptor, *e.g.*, agonists and antagonists of the modulators of the invention in mammals.
- 20 It is a still further object of the present invention to provide a method for the treatment of mammals to control body weight and fat content in mammals, and/or to treat certain of the pathological conditions of which abnormal depression or elevation of body weight is a characterizing feature.
- 25 It is a still further object of the present invention to prepare genetic constructs for use in genetic therapeutic protocols and/or pharmaceutical compositions for comparable therapeutic methods, which comprise or are based upon one or more of the modulators, binding partners, or agents that may control their production,
- 30 or that may mimic or antagonize their activities.

Other objects and advantages will become apparent to those skilled in the art from a review of the ensuing description which proceeds with reference to the following illustrative drawings.

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BRIEF DESCRIPTION OF THE DRAWINGS

FIGURE 1. *Localization of the leptin receptor to the region of the db gene.*

The *db* mutation was segregated in two crosses totaling 750 meioses. A genetic map was compiled by genotyping the progeny of these crosses with the markers indicated in the map. Key recombinant animals are noted on the map as numbers above the line. A chromosome walk was initiated with the microdissection clone D4Rck22. The walk spanned 2.7 megabases and was composed of YACs (bold lines), BACs (italics) and P1 bacteriophage (numbers). Genotyping of the recombinant animals with two SSLP markers, D4Rck6 and D4Rck7 from the ends of these genomic clones, localized the *db* gene to the approximately 300 Kb interval between the recombination events in animals 324 and 1028. This interval was spanned by BACs 242 and 43. Southern blots and PCR revealed that the 5' ends of the leptin receptor mapped to BAC 150A and the 3' end to BAC 19, indicating the gene is transcribed toward the telomeres.

FIGURE 2. *Several splice variants of the leptin receptor are present.* (A) A schematic drawing of the leptin receptor, with putative motifs for JAK binding and signal transduction, "Box 1" and "Box 2" (shaded areas). "TM" indicates a putative transmembrane domain. A total of 8 cDNA clones were isolated from mouse brain. These cDNAs were found to correspond to five different splice variants of the leptin receptor. (B) Six of the clones had partly identical sequences upstream of lysine 889 of the leptin receptor (OB-Ra, OB-Rb, OB-Rc and OB-Rd), at which point the predicted proteins diverged. The predicted C-terminal amino acid sequences of these clones is shown (SEQ ID NOS: 11-14, respectively). OB-Ra, b, c, and d all predict a Box 1 motif. OB-Rb also predicts a peptide sequence

potentially homologous to Box 2 (underlined). Two independent cDNA clones were identical to the leptin receptor upstream of histidine 796, at which point the sequences diverged (OB-Re)(SEQ ID NO:15). The nucleotide sequence predicts a soluble receptor.

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FIGURE 3. *The db mutation results in abnormal RNA splicing and conversion of the splice variant OB-Rb to OB-Ra.* (A) RT-PCR products from C57BL/Ks db/db and wild type mice were amplified using a primer pair specific for OB-Rb RNA (F1 and R₃). Electrophoresis revealed that the amplified fragment from these db mice was larger than from wild type animals. The PCR products of genomic DNA spanning the OB-Rb splice acceptor at Pro⁸⁹⁰ were of identical size in C57 BL/Ks db/db mice and littermate controls. (B) Primers F2 and R were used to amplify the genomic DNA. The F2 primer was selected after using vectorette PCR and BAC 242 to obtain the sequence of genomic DNA upstream of the splice acceptor at P⁸⁹⁰. (C) Localization of primers for RT-PCR and genomic PCR amplification.

FIGURE 4. *Hypothalamic RNA of wild type mice.* The hypothalamic RT-PCR products for the C-terminal coding region of (A) OB-Ra, (B) OB-Re, (C) OB-Rd and (D) OB-Re were of normal size in db mice. The DNA sequence across the splice junction was normal in each of these RT-PCR products. This indicates that the splice donor at Lys⁸⁸⁹ is wild type.

FIGURE 5. *Identification of splice mutations in db mice.* (A) DNA sequencing identified a 106 bp insertion in the mutant OB-Rb RNA at the splice junction between Lys⁸⁸⁹ and Pro⁸⁹⁰ (SEQ ID NO:16). The sequence of the insertion was identical to the first 106 bp of the C-terminal exon of OB-RA. The insertion predicts a premature stop codon and changes the amino acid sequence of OB-Rb (SEQ ID NO:17) to OB-Ra. (B, C) The presumed genomic organization of the OB-Ra and OB-Rb 3' ends are shown. DNA sequencing of the OB-Ra exon from the C57 BL/K⁹ db/db mice (SEQ ID NO: 18) and littermate controls (SEQ ID

NO:19) revealed a G to T mutation 106 base pairs after the splice acceptor at R⁸⁹⁰. This mutation results in the appearance of a consensus splice donor site, AGGTAAA, which leads to the insertion of 106 bp of the C-terminal exon of OB-Ra into that of OB-Rb.

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FIGURE 6. *Tissue distribution of the alternatively spliced leptin receptor.*

RT-PCR was performed from the tissue sources indicated. In each case, one primer from a region of shared nucleotide sequence was used in combination with a primer specific for the alternatively spliced exon. (B) Brain, (H) Hypothalamus,
 10 (L) Liver, (H) Heart, (K) Kidney, (S) Spleen, (T) Testis, (F) Adipose Tissue, (S) Spleen.

DETAILED DESCRIPTION OF THE INVENTION

15 The present invention relates to the elucidation and discovery of a protein, termed herein ob receptor (OB-R) or leptin receptor, nucleic acids encoding the protein, including the *OB-R* gene (also termed herein *DB* -- it should be noted that where all capitals are used it refers to the natural protein or gene; all lower case refers to a mutant protein or gene; italics indicates a gene or nucleic acid molecule; and
 20 normal type indicates a protein or polypeptide), including degenerate variations thereof, *e.g.*, that incorporate optimal codons for expression in a particular expression system, which protein demonstrates the ability to participate in the control of mammalian body weight. In particular, the protein demonstrates the ability to bind leptin. In a specific embodiment, the protein mediates signal
 25 transduction upon binding to leptin.

The OB receptor of the invention may contain three important structural domains: an extracellular (or extracytoplasmic) domain, a transmembrane domain, and a cytoplasmic domain. The extracellular domain is postulated to bind leptin, leptin-
 30 protein complexes (such as leptin bound to a soluble leptin receptor), and may possibly bind other proteins or ligands. In a specific embodiment, a receptor of

the invention comprises only an extracellular domain, *i.e.*, it is a soluble receptor. The transmembrane domain comprises a stretch of highly non-polar amino acid residues that localize to the hydrophobic region of the cell membrane. In this respect, the term transmembrane domain has its ordinary meaning in molecular biology. Finally, the cytoplasmic domain of an OB receptor of the invention may contain none, one, or two JAK-binding consensus sequences, termed "Box 1" and "Box 2". A receptor having "Box 1" and "Box 2" is believed competent for signal transduction via the JAK-Stat pathway upon bind ligand, *e.g.*, leptin.

- 10 Furthermore, the protein has been identified as having numerous splice-forms. In one aspect, the splice variations lead to divergence of the C-terminal sequences. Thus, the protein can be found in a secreted form postulated to agonize leptin activity; it can be found as an integral membrane receptor that may facilitate leptin transfer across the blood-brain barrier, but that lacks domains involved in signal transduction; and it can be found as a integral membrane receptor containing domains involved in signal transduction. In another aspect, splice variations lead to divergence of the N-terminal polypeptide sequence.

- 20 The nucleic acids in object represent the coding sequences corresponding to the animal, specifically murine and human OB-R polypeptide, which, by mediating (or failing to mediate) signal transduction on binding leptin, is postulated to play a critical role in the regulation of body weight and adiposity. Data presented herein indicate that one splice variant of the polypeptide product of a nucleic acid of the invention may be secreted by the cells that express it, or it may be expressed as an integral membrane protein. In either event, the polypeptide functions as a leptin receptor. Additional experimental data suggest that the naturally occurring splice-form of the OB-R polypeptide is very effective in treating obesity in mice carrying a mutation of the *ob* gene.

- 30 In addition, the Examples herein demonstrate that mRNA encoding the OB-R polypeptide, alternatively termed herein "leptin receptor," is expressed in

hypothalamus, testes, and adipocytes. Data also demonstrate expression of the protein in the choroid plexus.

In a further aspect, the OB-R polypeptide from one species is closely related
 5 (homologous) to the OB-R in another species. In particular, the human OB-R polypeptide is highly homologous to murine OB-R polypeptide. This observation is consistent with the data showing that human leptin is active in mice: for the hormone to be active ~~interspecies~~ ^{interspecies}, one would expect a high degree of similarity or homology between the receptors from different species as well.

10

In its primary aspect, the present invention is directed to the identification of materials that function as modulators of mammalian body weight. In particular, the invention concerns the isolation, purification, and sequencing of certain nucleic acids that correspond to the *OB-R* gene (alternatively referred to herein and in the
 15 literature as *DB*) or its coding region in both mice and humans, as well as the corresponding polypeptides expressed by these nucleic acids. The invention thus comprises the discovery of nucleic acids having the nucleotide sequences set forth in SEQ ID NOS: ~~1-5~~ ^{1, 3, 5, 7 and 9}, and to degenerate variants, alleles and fragments thereof, all possessing the activity of modulating body weight and adiposity. The
 20 correspondence of the present nucleic acids to the *OB-R* gene portends their significant impact on conditions such as obesity as well as other maladies and dysfunctions where abnormalities in body weight are a contributory factor. The invention extends to the proteins expressed by the nucleic acids of the invention, and particularly to those proteins set forth in SEQ ID NOS: ~~6-10~~ ^{2, 4, 6, 8 and 10}, as well as to
 25 conserved variants and active fragments.

Of particular interest according to the invention are different splice variants of OB-R, e.g., as represented by OB-Ra, OB-Rb, OB-Rc, OB-Rd, and OB-Re. The present invention anticipates other OB-R splice variants as well.

30

Thus, in specific embodiments, the term OB-R refers to splice variants as follows (amino acid numbering correspond to the numbering applied to murine OB-R [Tartaglia *et al.*, *Cell*, 83:1263 (1995)], which has been adopted herein):

- 5 N-terminal corresponding to OB-Ra through Lys⁸⁸⁹ and C-terminal corresponding to a C-terminal selected from the group consisting of OB-Rb, OB-Rc, and OB-Rd after Lys⁸⁸⁹;

 N-terminal corresponding to OB-Rb or OB-Rc through Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra or OB-Rd after Lys⁸⁸⁹;

- 10 N-terminal corresponding to OB-Rd through Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra, OB-Rb, or OB-Rc;

 N-terminal corresponding to OB-R from Pro⁶⁶⁴ to Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra, OB-Rb, OB-Rc, and OB-Rd;

- 15 N-terminal corresponding to OB-R from Met⁷³³ to Lys⁸⁸⁹, and C-terminal corresponding to OB-Ra, OB-Rb, OB-Rc, and OB-Rd;

 N-terminal selected from the group consisting of OB-Ra, OB-Rb, OB-Rd, and OB-R from Pro⁶⁶⁴, to His⁷⁹⁶, and OB-Re from His⁷⁹⁶; and

 N-terminal corresponding to OB-R from Met⁷³³ to His⁷⁹⁶, and OB-Re from His⁷⁹⁶.

20

Various forms of the OB-R, which may act as agonists (*e.g.*, the naturally occurring secreted form of the OB-R) or antagonists (*e.g.*, a truncated form of OB-R that only binds leptin), may be prepared in pharmaceutical compositions, with a suitable carrier and at a strength effective for administration by various

- 25 means to a patient experiencing abnormal fluctuations in body weight or adiposity, either alone or as part of an adverse medical condition such as cancer or AIDS, for the treatment thereof. A variety of administrative techniques may be utilized, among them oral administration, nasal and other forms of transmucosal administration, parenteral techniques such as subcutaneous, intravenous and
- 30 intraperitoneal injections, catheterizations and the like. Appropriate quantities of

the soluble OB-R molecules may vary and in particular should be based upon the recommendations and prescription of a qualified physician or veterinarian.

In accordance with the above, an assay system for screening potential drugs
5 effective to mimic or antagonize the activity of leptin may be prepared. The
prospective drug may be contacted with a soluble form of the OB-R, or
alternatively may be used with cells that express a receptor form of OB-R, to
determine whether it binds to, or activates (or antagonizes) OB-R. For example,
in an expression assay system, the culture may be examined to observe any
10 changes in the activity of the cells, due either to the addition of the prospective
drug alone, or due to the effect of added quantities of the known weight
modulator.

As stated earlier, the molecular cloning of the *OB-R* gene described herein has led
15 to the identification of a class of materials that function on the molecular level to
modulate mammalian body weight. The discovery of the modulators of the
invention has important implications for the diagnosis and treatment of nutritional
disorders including, but not limited to, obesity, weight loss associated with cancer
and the treatment of diseases associated with obesity such as hypertension, heart
20 disease, and Type II diabetes. In addition, there are potential agricultural uses for
the gene product in cases where one might wish to modulate the body weight of
domestic animals. The discussion that follows with specific reference to the *OB-R*
gene bears general applicability to the class of modulators that comprise a part of
the present invention, and is therefore to be accorded such latitude and scope of
25 interpretation.

In a particular embodiment, the functional activity of the OB-R polypeptide can be
evaluated transgenically. The *OB-R* gene can be used in complementation studies
employing transgenic mice. Transgenic vectors, including viral vectors, or cosmid
30 clones (or phage clones) corresponding to the wild type locus of candidate gene,
can be constructed using the isolated *OB-R* gene. Cosmids may be introduced into

transgenic mice using published procedures [Jaenisch, *Science*, **240**:1468-1474 (1988)]. The constructs are introduced into fertilized eggs derived from an intercross between F1 progeny of a C57BL/6J *db/db* X DBA intercross. Genotype at the *db* loci in cosmid transgenic animals can be determined by typing animals

5 with tightly linked RFLPs or microsatellites which flank the mutation and which are polymorphic between the progenitor strains. Complementation will be demonstrated when a particular construct renders a genetically obese F2 animal (as scored by RFLP analysis) lean and nondiabetic. Under these circumstances, final proof of complementation will require that the *db/db* animal carrying the transgene

10 be mated to the *db/db* ovarian transplants. In this cross, all N2 animals which do not carry the transgene will be obese and insulin resistant/diabetic, while those that do carry the transgene will be lean and have normal glucose and insulin concentrations in plasma. In a genetic sense, the transgene acts as a suppressor mutation.

15 Alternatively, *OB-R* genes can be tested by examining their phenotypic effects when expressed in antisense orientation in wild-type animals. In this approach, expression of the wild-type allele is suppressed, which leads to a mutant phenotype. RNA-RNA duplex formation (antisense-sense) prevents normal

20 handling of mRNA, resulting in partial or complete elimination of wild-type gene effect. This technique has been used to inhibit TK synthesis in tissue culture and to produce phenotypes of the *Kruppel* mutation in *Drosophila*, and the *Shiverer* mutation in mice Izant *et al.*, *Cell*, **36**:1007-1015 (1984); Green *et al.*, *Annu. Rev. Biochem.*, **55**:569-597 (1986); Katsuki *et al.*, *Science*, **241**:593-595 (1988). An

25 important advantage of this approach is that only a small portion of the gene need be expressed for effective inhibition of expression of the entire cognate mRNA. The antisense transgene will be placed under control of its own promoter or another promoter expressed in the correct cell type, and placed upstream of the SV40 polyA site. This transgene can be used to make transgenic mice.

30 Transgenic mice can also be mated ovarian transplants to test whether *ob* heterozygotes are more sensitive to the effects of the antisense construct.

In the long term, the *OB-R* gene product (the OB-R polypeptide or protein) is useful for identifying small molecule agonists and antagonists that affect its activity.

- 5 Various terms used throughout this specification shall have the definitions set out herein, for example, below.

The term "body weight modulator", "modulator", "modulators", and any variants not specifically listed, may be used herein interchangeably, and as used throughout
10 the present application and claims refers in one instance to both nucleotides and to proteinaceous material, the latter including both single or multiple proteins. More specifically, the aforementioned terms extend to the nucleotides and to the DNA having the sequences described herein and presented in SEQ ID NOS:1, 3, 5, 7, and 9. Likewise, the proteins having the amino acid sequence data described
15 herein and presented in SEQ ID NOS: 2, 4, 6, 8, and 10 are likewise contemplated, as are the profile of activities set forth with respect to all materials both herein and in the claims.

Specific binding to leptin means that leptin is a ligand for OB-R, as that term is
20 used to describe ligand-receptor binding. Generally, such binding will have an affinity represented by an association constant of greater than $1 \times 10^7 \text{M}^{-1}$, preferably greater than $1 \times 10^8 \text{M}^{-1}$, and more preferably greater than $1 \times 10^9 \text{M}^{-1}$. However, the exact association constant may vary. Homology with gp130 refers to conservation of residues, particularly cysteine residues, motifs, and other
25 important residues. The term "gp130" is used herein to refer generally to the class I cytokine receptor family, particularly interleukin-6 (IL-6) receptor, granulocyte colony-stimulating factor (G-CSF) receptor, ciliary neurotrophic factor (CNTF) receptor, and leukemia inhibitory factor (LIF) receptor.

- 30 Additionally, nucleotides displaying substantially equivalent or altered activity are likewise contemplated, including substantially homologous analogs and allelic

variations. Likewise, proteins displaying substantially equivalent or altered activity, including proteins modified deliberately, as for example, by site-directed mutagenesis, or accidentally through mutations in hosts that produce the modulators are likewise contemplated.

5

The term "allelic variants" refers to the corresponding gene in different individuals that may have point mutations. For example, the various *ob* mutation represent allelic variants of *OB-R*.

- 10 The term "substantially homologous analogs" specifically includes the corresponding gene or protein from another species. In a specific embodiment, a substantially homologous analog of murine OB-R is human OB-R. The term can also include genes or proteins mutated or altered, *e.g.*, by substitution of variant amino acid residues from one species in the polypeptide of another, so as to
- 15 correspond to an analogous gene or protein as if from another species.

The term "gene" as used herein refers to a nucleic acid, such as DNA, which codes on expression for a protein. Unless stated otherwise, gene may include mRNA, cDNA, or genomic DNA.

20

- A composition comprising "A" (where "A" is a single protein, DNA molecule, vector, recombinant host cell, etc.) is substantially free of "B" (where "B" comprises one or more contaminating proteins, DNA molecules, vectors, etc., but excluding racemic forms of A) when at least about 75% by weight of the proteins, DNA, vectors (depending on the category of species to which A and B belong) in
- 25 the composition is "A". Preferably, "A" comprises at least about 90% by weight of the A+B species in the composition, most preferably at least about 99% by weight. It is also preferred that a composition, which is substantially free of contamination, contain only a single molecular weight species having the activity
- 30 or characteristic of the species of interest.

A "BAC" is a bacterial artificial chromosome; "STS" refers to sequence tagged site; a "YAC" is a yeast artificial chromosome. Other terms have the standard meanings ordinarily intended in the art.

5

The OB-R Polypeptides

The terms "protein," which refers to the naturally occurring polypeptide, and "polypeptide" are used herein interchangeably with respect to the *OB-R* gene product and variants thereof. More particularly, OB-R refers to any of the splice forms of the *OB-R (DB)* gene product, such as but not limited to the product with two JAC binding boxes in the cytoplasmic domain; the product with only one JAK binding box in the cytoplasmic domain; the product with no boxes; and the secreted (soluble) product. The term OB-R also refers to various splice-forms with divergent N-terminal amino acid sequences.

The term OB-R specifically encompasses different splice forms of the polypeptide, including but not limited to the follows:

<u>Splice Form</u>	<u>Characteristics</u>	<u>Specific Embodiment</u>
OB-Ra	Transmembrane protein with a "Box 1" but no "Box 2"; expected to bind leptin but does not directly mediate signal transduction via JAKs. Comprised of an extracellular domain, and a truncated cytoplasmic domain. N-terminus diverges from published OB-R sequence upstream of Cys ⁸⁸ .	SEQ ID NO:2
OB-Rb	Transmembrane protein expected to mediate leptin signalling in hypothalamus and other cells. contains a larger cytoplasmic domain containing both a "Box 1" and "Box 2" sites. N-terminal	SEQ ID NO:4

portion appears to be truncated, diverging from the published OB-R sequence upstream of Pro⁶⁶⁴.

- | | | | |
|----|-------|--|--------------|
| 5 | OB-Rc | Corresponds to OB-Rb with a tripeptide residue C-terminal to Lys ⁸⁸⁹ rather than the longer sequence; no "Box 2" site. | SEQ ID NO:6 |
| 10 | OB-Rd | Corresponds to published OB-R with a different <u>eleven</u> amino acid sequence C-terminal to Lys ⁸⁸⁹ . | SEQ ID NO:8 |
| 15 | OB-Re | Soluble/secreted receptor with a leptin-binding domain. Lacks a transmembrane or cytoplasmic domain, but comprises a large extracellular domain. | SEQ ID NO:10 |
| 20 | | Corresponds to published OB-R to His ⁷⁹⁶ , where it diverges . | |

The term OB-R specifically contemplates splice variants that incorporate different elements from the above-noted variants, *e.g.*, as described above.

More particularly, the present invention is directed to OB-R with the N-terminal signal sequence cleaved. In one embodiment, amino acid residues 1-22 are cleaved. In another embodiment, amino acid residues 1-27 are cleaved.

As noted above, in specific embodiments polypeptides of the invention include those having the amino acid sequences set forth herein *e.g.*, SEQ ID NOS:2, 4, 6, 8, and 10. The term further includes polypeptides modified with conservative amino acid substitutions, as well as biologically active fragments, analogs, and derivatives thereof. The term "biologically active," is used herein to refer to a specific effect of the polypeptide, including but not limited to specific binding, *e.g.*, to leptin, an anti-OB-R antibody, or other recognition molecule; activation of signal transduction pathways on a molecular level; and/or induction (or inhibition by antagonists) of physiological effects mediated by the native leptin *in vivo*. OB-

R polypeptides, including fragments, analogs, and derivatives, can be prepared synthetically, *e.g.*, using the well known techniques of solid phase or solution phase peptide synthesis. Preferably, solid phase synthetic techniques are employed. Alternatively, OB-R polypeptides of the invention can be prepared
5 using well known genetic engineering techniques, as described *infra*. In yet another embodiment, the soluble form of the OB-R polypeptide can be purified, *e.g.*, by immunoaffinity purification, from a biological fluid, such as but not limited to plasma, serum, or urine, preferably human plasma, serum, or urine, and more preferably from a subject who overexpresses the polypeptide.

10

The structure of the OB-R polypeptide, preferably human OB-R polypeptide, can be analyzed by various methods known in the art. The protein sequence can be characterized by a hydrophilicity analysis [*e.g.*, Hopp *et al.*, *Proc. Natl. Acad. Sci. USA*, **78**:3824 (1981)]. A hydrophilicity profile can be used to identify the
15 hydrophobic and hydrophilic regions of the OB-R polypeptide, which may indicate regions buried in the interior of the folded polypeptide, the transmembrane domain, and regions accessible on the exterior of the polypeptide. In addition, secondary structural analysis [*e.g.*, Chou *et al.*, *Biochem.*, **13**:222 (1974)] can also be done, to identify regions of OB-R polypeptide that assume specific secondary
20 structures. Manipulation of the predicted or determined structure, including secondary structure prediction, can be accomplished using computer software programs available in the art.

By providing an abundant source of recombinant OB-R polypeptide, the present
25 invention enables quantitative structural determination of the polypeptide. In particular, enough material is provided for nuclear magnetic resonance (NMR), infrared (IR), Raman, and ultraviolet (UV), especially circular dichroism (CD), spectroscopic analysis. In particular NMR provides very powerful structural analysis of molecules in solution, which more closely approximates their native
30 environment [Marion *et al.*, *Biochim. Biophys. Res. Comm.*, **113**:967-974 (1983); Bar *et al.*, *J. Magn. Reson.*, **65**:355-360 (1985); Kimura *et al.*, *Proc. Natl. Acad.*

Sci. USA, 77:1681-1685 (1980)]. Other methods of structural analysis can also be employed. These include but are not limited to X-ray crystallography [Engstrom, *Biochem. Exp. Biol.*, 11:7-13 (1974)]. In a preferred aspect, either soluble form or a membrane-binding form of OB-R is co-crystallized with leptin to provide
 5 structural information about both molecules.

In yet a further embodiment, an analog of OB-R polypeptide can be tested to determine whether it cross-reacts with an antibody specific for native OB-R polypeptide, or specific fragments thereof. The degree of cross-reactivity provides
 10 information about structural homology or similarity of proteins, or about the accessibility of regions corresponding to portions of the polypeptide that were used to generate fragment-specific antibodies.

Fragments of the OB-R Polypeptide

15 In a particular embodiment, the present invention contemplates that naturally occurring fragments, or truncated forms, of the OB-R polypeptide may be important. As noted above, a large number of splice forms of OB-R have been found. Thus, the present invention encompasses a naturally occurring soluble form of the OB-R, as well as integral membrane forms that have 0, 1, or 2 JAK
 20 box consensus sites. In addition to the naturally occurring splice isoforms of the polypeptide, the present invention further envisions recombinantly modified isoforms, *e.g.*, by deletion of one or more of the cytoplasmic domain; the cytoplasmic consensus domain from the transmembrane domain to lysine-889; the box 1 or box two, or both regions; the cytoplasmic domain C-terminal of lysine-
 25 889; the transmembrane domain; the ligand binding domain; the extracytoplasmic domain; or portions thereof.

OB-R Polypeptide Chimeras

One or more of the splice-forms of the cytoplasmic domain can be used in a
 30 chimeric construct with another ligand-binding domain to artificially signal leptin binding [*e.g.*, Capon *et al.*, U.S. Patent No. 5,359,046, issued October 25, 1994;

Sanchez *et al.*, *J. Exp. Med.*, **178**:1049 (1993); Burkhardt *et al.*, *Mol. Cell. Biol.*, **14**:1095]. In another embodiment, the extracytoplasmic (leptin-binding) domain can be joined to a different cytoplasmic signal transduction domain, or alternatively to a glycosyl-phosphatidylinositol linker domain to provide for
 5 activation of cells via gp130.

Analogs of the OB-R Polypeptide

The present invention specifically contemplates preparation of analogs of the OB-R polypeptide, which are characterized by being capable of a biological activity of
 10 OB-R polypeptide, *e.g.*, of binding to leptin or to an anti-OB-R antibody. In one embodiment, the analog agonizes OB-R activity. Preferably, an OB-R agonist is more effective than the native protein. For example, an OB-R agonist analog may bind to leptin with higher affinity, thus amplifying the signal. Such an analog may be particularly desirable for gene therapy, where increased signal transduction
 15 efficiency can compensate for any deficiency in the level of expression. In another embodiment, the analog antagonizes OB-R activity. For example, an OB-R analog that binds to leptin, and inhibits leptin binding to signal-transduction competent OB-R, can competitively inhibit binding of native OB to the receptor, thus decreasing leptin activity *in vivo*. Such an OB-R antagonist analog is
 20 preferably a soluble form of the OB-R.

In one embodiment, an analog of OB-R polypeptide is the OB-R polypeptide modified by substitution of amino acids at positions on the polypeptide that are not essential for structure or function. For example, since it is expected that human
 25 OB-R polypeptide is biologically active in mouse, substitution of divergent amino acid residues in the human sequence as compared to the murine amino acid sequence will likely yield useful analogs of OB-R polypeptide. For example, the following residues in the human OB-R [numbering for human OB-R amino acids employs the numbering convention adopted in Tartaglia *et al.*, *Cell*, **83**:1263
 30 (1995)] could be substituted with a divergent murine residue found at that position, or with a non-conservative amino acid substitution, such as one or more of: Phe

for Ser³⁶; Asp for Tyr⁴⁴; Ser for Leu⁴⁹; Pro for Ser⁵⁴; Leu for Ser⁶⁰; Ala for His⁶³; Ala for Thr⁶⁶; Ala for Pro⁷⁰; Ile for Thr⁷⁷; Tyr for His⁷⁸; Pro for Ser⁸⁰; Gly for Arg⁹²; Gly for Asp⁹⁶; Thr for Ala¹⁰³ or Ile¹⁰⁶; Ser for Leu¹¹⁸; Gly for Asp¹²⁴; Thr for Lys¹³⁸; Pro for Ser¹⁴⁶; Asp for Val¹⁶⁴; Leu for Gln¹⁷⁷; Asp for Gly¹⁷⁹; Gly for Glu¹⁹²; deletion for Cys¹⁹³; His for Leu¹⁹⁷; Ser for Ile²²¹; Leu for Asn²³³; Leu for Ser²⁷³; deletion for Thr²⁷⁸; Ala for Asp²⁸⁵; Glu for Lys²⁸⁶; Ser for Gly³¹⁰; Arg for Met³⁷⁰; Ile for Ser³⁷⁹; Ser for Phe³⁹⁴; Ala for Glu⁴¹⁷; Gly for Glu⁴⁵⁹; Ser for Ile⁴⁷⁶; Thr for Ile⁴⁸²; Thr for Ile⁵⁵¹; His for Tyr⁵⁸⁶; Lys for Ile⁶⁴⁸; Ala for Ser⁶⁸⁶; His for Cys⁶⁸⁷; Thr for Ile⁷⁵⁹; Ile for Asn⁷⁷⁶; Asp for Gly⁷⁸¹; Gly for Glu⁷⁸²; Gly for Ser⁸²⁷; Ala for Asp⁸³²; Arg for Pro⁸⁹²; Thr for Glu⁸⁹³; Asp for Thr⁸⁹⁴; or Leu for Glu⁸⁹⁶.

Also contemplated by the present invention are analogs comprising conservative amino acid substitutions. For example, one or more amino acid residues within the sequence can be substituted by another amino acid of a similar polarity, which acts as a functional equivalent, resulting in a silent alteration. Substitutes for an amino acid within the sequence may be selected from other members of the class to which the amino acid belongs. For example, the nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan and methionine. The polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine. The positively charged (basic) amino acids include arginine, lysine and histidine. The negatively charged (acidic) amino acids include aspartic acid and glutamic acid. In some instances, one polar amino acid may be substituted with another to preserve local hydrophilicity; more likely, a substitution that conserves charge, or at least does not introduce the opposite charge, is required. Such alterations will not be expected to affect apparent molecular weight as determined by polyacrylamide gel electrophoresis, or isoelectric point.

In still another embodiment, amino acid residues can be substituted with residues to form analogs of OB-R polypeptide that demonstrate enhanced propensity for forming, or which form more stable, secondary structures. For example, α -helix

structure would be preferred if Glu, Ala, Leu, His, Trp are introduced as substitutes for amino acid residues found in the native OB polypeptide.

Preferably, conservative amino acid substitutions are employed, *e.g.*, substituting aspartic acid with glutamic acid(s) (Glu); substituting isoleucine(s) with leucine; substituting glycine or valine, or any divergent amino acid (*i.e.*, an amino acid that is not conserved between OB-R from different species), with alanine (*e.g.*, serine at position 273 of the human OB-R polypeptide with alanine); substituting arginine or lysine with histidine; and substituting tyrosine and/or phenylalanine with tryptophan. Increasing the degree, or more importantly, the stability of α -helix structure may yield an OB-R analog with greater activity, increased binding affinity, or longer half-life. Also contemplated are truncated OB-R polypeptide analogs are generated that incorporate structure-forming, *e.g.*, helix-forming, amino acid residues to compensate for the greater propensity of polypeptide fragments to lack stable structure.

15

In another embodiment, an analog of the OB-R polypeptide, preferably the human OB-R polypeptide, is a truncated form of the polypeptide. For example, it has already been demonstrated that the transmembrane domain is not essential, since a naturally occurring isoform of the polypeptide is encoded by cDNA that expresses a soluble protein. Similarly, it may be possible to delete some or all of the divergent amino acid residues (as compared to the murine OB-R). In addition, the invention contemplates providing an OB-R analog having the minimum amino acid sequence necessary for a biological activity. This can be readily determined, *e.g.*, by testing the activity of fragments of OB-R for the ability to bind to OB-R-specific antibodies, inhibit the activity of the native leptin (by competitive binding), or agonize the activity of native leptin.

20

25

The present invention specifically contemplates providing a soluble splice-form of the OB-R that is believed to agonize leptin activity. In particular, it is believed that OB-Rd (as referred to herein) binds leptin, and facilitates leptin binding to OB-Re (which is believed to be competent for signal transduction). Thus, in this

30

embodiment, OB-R appears to behave analogously to other receptor systems [Kishimoto *et al.*, *Cell*, **76**:253 (1994); Davis *et al.*, *Science*, **260**:1805 (1993); Davis *et al.*, *Science*, **259**:1736 (1993)].

- 5 It will be appreciated by one of ordinary skill in the art that the foregoing fragment sizes are approximate, and that additional amino acids *e.g.* from one to about five, can be included or deleted from each or both ends, or from the interior of the polypeptide or fragments thereof, of the recited truncated analogs.
- 10 Analogs, such as fragments, may be produced, for example, by pepsin digestion of the OB-R. Other analogs, such as muteins, can be produced by standard site-directed mutagenesis of weight modulator peptide coding sequences.

Screening for Leptin Analogs

- 15 Various screening techniques are known in the art for screening for analogs of polypeptides. Various libraries of chemicals are available. Accordingly, the present invention contemplates screening such libraries, *e.g.*, libraries of synthetic compounds generated over years of research, libraries of natural compounds, and combinatorial libraries, as described in greater detail, *infra*, for analogs of leptin.
- 20 The invention contemplates screening such libraries for compounds that bind to OB-R, either in soluble or transmembrane forms. Preferably, such molecules agonize or antagonize signal transduction by OB-R. The present invention contemplates screens for small molecule ligands or ligand analogs and mimics, as well as screens for natural ligands that bind to and agonize or antagonize activate
- 25 OB receptor *in vivo*.

- Knowledge of the primary sequence of the receptor, and the similarity of that sequence with proteins of known function, can provide an initial clue as to the agonists or antagonists of the protein. Identification and screening of antagonists
- 30 is further facilitated by determining structural features of the protein, *e.g.*, using X-ray crystallography, neutron diffraction, nuclear magnetic resonance

spectrometry, and other techniques for structure determination. These techniques provide for the rational design or identification of agonists and antagonists.

Another approach uses recombinant bacteriophage to produce large libraries.

- 5 Using the "phage method" [Scott *et al.*, *Science*, **249**:386-390 (1990); Cwirla *et al.*, *Proc. Natl. Acad. Sci. USA*, **87**:6378-6382 (1990); Devlin *et al.*, *Science*, **249**:404-406 (1990)], very large libraries can be constructed (10^6 - 10^8 chemical entities). A second approach uses primarily chemical methods, of which the Geysen method [Geysen *et al.*, *Molecular Immunology*, **23**:709-715 (1986);
- 10 Geysen *et al.*, *J. Immunologic Method*, **102**:259-274 (1987)] and the recent method of Fodor *et al.*, *Science*, **251**:767-773 (1991) are examples. Furka *et al.* 14th International Congress of Biochemistry, Volume 5, Abstract FR:013 (1988); Furka, *Int. J. Peptide Protein Res.*, **37**:487-493 (1991)]; Houghton (U.S. Patent No. 4,631,211, issued December 1986); and Rutter *et al.* (U.S. Patent No.
- 15 5,010,175, issued April 23, 1991) describe methods to produce a mixture of peptides that can be tested as agonists or antagonists.

- In another aspect, synthetic libraries [Needels *et al.*, *Proc. Natl. Acad. Sci. USA*, **90**:10700-10704 (1993); Lam *et al.*, International Patent Publication No. WO
- 20 92/00252, each of which is incorporated herein by reference in its entirety], and the like can be used to screen for OB receptor ligands according to the present invention.

- In particular, assays for binding of soluble ligand to cells that express recombinant
- 25 forms of the OB receptor ligand binding domain can be performed. The soluble ligands can be provided readily as recombinant or synthetic leptin polypeptide.

- The screening can be performed with recombinant cells that express the OB receptor, or alternatively, using purified receptor protein, *e.g.*, produced
- 30 recombinantly, as described above. For example, the ability of labeled, soluble, or solubilized OB receptor, that includes the ligand-binding portion of the

molecule, to bind ligand can be used to screen libraries, as described in the foregoing references.

Derivatives of OB Polypeptides

- 5 Generally, a soluble form of the present polypeptide may be derivatized by the attachment of one or more chemical moieties to the polypeptide moiety. The chemically modified derivatives may be further formulated for intraarterial, intraperitoneal, intramuscular, subcutaneous, intravenous, oral, nasal, rectal, bucal, sublingual, pulmonary, topical, transdermal, or other routes of
- 10 administration. Chemical modification of biologically active proteins has been found to provide additional advantages under certain circumstances, such as increasing the stability and circulation time of the therapeutic protein and decreasing immunogenicity. See U.S. Patent No. 4,179,337, Davis *et al.*, issued December 18, 1979. For a review, see Abuchowski *et al.*, "Soluble
- 15 Polymer-Enzyme Adducts", in *Enzymes as Drugs*, pp. 367-383, Holcenberg and Roberts, eds., Wiley-Interscience, New York, NY, (1981). A review article describing protein modification and fusion proteins is Francis, *Focus on Growth Factors*, 3:4-10 (1992).

Chemical Moieties For Derivatization

- 20 The chemical moieties suitable for derivatization may be selected from among water soluble polymers. The polymer selected should be water soluble so that the protein to which it is attached does not precipitate in an aqueous environment, such as a physiological environment. Preferably, for therapeutic use of the
- 25 end-product preparation, the polymer will be pharmaceutically acceptable. One skilled in the art will be able to select the desired polymer based on such considerations as whether the polymer/protein conjugate will be used therapeutically, and if so, the desired dosage, circulation time, resistance to proteolysis, and other considerations. For the present proteins and peptides, these
- 30 may be ascertained using the assays provided herein.

Polymer Molecules

The water soluble polymer may be selected from the group consisting of, for example, polyethylene glycol, copolymers of ethylene glycol/propylene glycol, carboxymethylcellulose, dextran, polyvinyl alcohol, polyvinyl pyrrolidone, poly-1,
5 3-dioxolane, poly-1,3,6-trioxane, ethylene/maleic anhydride copolymer, polyaminoacids (either homopolymers or random copolymers), and dextran or poly(n-vinyl pyrrolidone)polyethylene glycol, propylene glycol homopolymers, polypropylene oxide/ethylene oxide co-polymers, polyoxyethylated polyols and polyvinyl alcohol. Polyethylene glycol propionaldehyde may provide advantages
10 in manufacturing due to its stability in water.

The polymer may be of any molecular weight, and may be branched or unbranched. For polyethylene glycol, the preferred molecular weight is between about 2kDa and about 100kDa (the term "about" indicating that in preparations of
15 polyethylene glycol, some molecules will weigh more, some less, than the stated molecular weight) for ease in handling and manufacturing. Other sizes may be used, depending on the desired therapeutic profile (*e.g.*, the duration of sustained release desired, the effects, if any on biological activity, the ease in handling, the degree or lack of antigenicity and other known effects of the polyethylene glycol
20 to a therapeutic protein or analog).

Polymer/Protein Ratio

The number of polypeptide molecules attached to each polymer may vary, and one skilled in the art will be able to ascertain the effect on function. One may mono-
25 derivatize, or may provide for a di-, tri-, tetra- or some combination of derivatization, with the same or different chemical moieties (*e.g.*, polymers, such as different weights of polyethylene glycols). The proportion of polymer molecules to protein (or peptide) molecules will vary, as will their concentrations in the reaction mixture. In general, the optimum ratio (in terms of efficiency of reaction
30 in that there is no excess unreacted protein or polymer) will be determined by factors such as the desired degree of derivatization (*e.g.*, mono, di-, tri-, etc.), the

molecular weight of the polymer selected, whether the polymer is branched or unbranched, and the reaction conditions.

Attachment of the Chemical Moiety to the Protein

- 5 The polyethylene glycol molecules (or other chemical moieties) should be attached to the protein with consideration of effects on functional or antigenic domains of the protein. There are a number of attachment methods available to those skilled in the art, *e.g.*, EP 0 401 384 herein incorporated by reference (coupling PEG to G-CSF). *See also* Malik *et al.*, *Exp. Hematol.*, **20**:1028-1035 (1992) (reporting
- 10 pegylation of GM-CSF using tresyl chloride). For example, polyethylene glycol may be covalently bound through amino acid residues via a reactive group, such as a free amino or carboxyl group. Reactive groups are those to which an activated polyethylene glycol molecule may be bound. The amino acid residues having a free amino group may include lysine residues and the N-terminal amino acid
- 15 residues; those having a free carboxyl group may include aspartic acid residues glutamic acid residues and the C-terminal amino acid residue. Sulfhydryl groups may also be used as a reactive group for attaching the polyethylene glycol molecule(s). Preferred for therapeutic purposes is attachment at an amino group, such as attachment at the N-terminus or lysine group. Attachment at residues
- 20 important for receptor binding should be avoided if receptor binding is desired.

N-terminally Chemically Modified Proteins.

- One may specifically desire N-terminally chemically modified protein. Using polyethylene glycol as an illustration of the present compositions, one may select
- 25 from a variety of polyethylene glycol molecules (by molecular weight, branching, etc.), the proportion of polyethylene glycol molecules to protein (or peptide) molecules in the reaction mix, the type of pegylation reaction to be performed, and the method of obtaining the selected N-terminally pegylated protein. The method of obtaining the N-terminally pegylated preparation (*i.e.*, separating this
- 30 moiety from other monopegylated moieties if necessary) may be by purification of the N-terminally pegylated material from a population of pegylated protein

molecules. Selective N-terminal chemical modification may be accomplished by reductive alkylation which exploits differential reactivity of different types of primary amino groups (lysine versus the N-terminus) available for derivatization in a particular protein. Under the appropriate reaction conditions, substantially

5 selective derivatization of the protein at the N-terminus with a carbonyl group containing polymer is achieved. For example, one may selectively N-terminally pegylate the protein by performing the reaction at a pH which allows one to take advantage of the pK_a differences between the ϵ -amino groups of the lysine residues and that of the α -amino group of the N-terminal residue of the protein. By such

10 selective derivatization attachment of a water soluble polymer to a protein is controlled: the conjugation with the polymer takes place predominantly at the N-terminus of the protein and no significant modification of other reactive groups, such as the lysine side chain amino groups, occurs. Using reductive alkylation, the water soluble polymer may be of the type described above, and should have a

15 single reactive aldehyde for coupling to the protein. Polyethylene glycol propionaldehyde, containing a single reactive aldehyde, may be used.

Nucleic Acids Associated With OB-R Polypeptide

20 As noted above, the present invention is directed to nucleic acids encoding OB-R polypeptides, as well as associated genomic non-coding sequences 5', 3', and intronic to the *OB-R* gene. Thus, in accordance with the present invention there may be employed conventional molecular biology, microbiology, and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in

25 the literature. See, e.g., Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1989); Glover ed., *DNA Cloning: A Practical Approach*, Volumes I and II, MRL Press, Ltd., Oxford, U.K. (1985); Gait ed., *Oligonucleotide Synthesis*, Oxford University Press (1984); Hames *et al.*, eds.,

30 *Nucleic Acid Hybridization*, Springer-Verlag (1985); Hames *et al.*, eds., *Transcription And Translation*, Oxford University Press (1984)]; Freshney ed.,

Animal Cell Culture, Oxford University Press (1986)]; *Immobilized Cells And Enzymes*, IRL Press (1986)]; Perbal, *A Practical Guide To Molecular Cloning*, Wiley, New York (1984). Of particular relevance to the present invention are strategies for isolating, cloning, sequencing, analyzing, and characterizing a gene
5 or nucleic acid based on the well known polymerase chain reaction (PCR) techniques.

A "replicon" is any genetic element (*e.g.*, plasmid, chromosome, virus) that functions as an autonomous unit of DNA replication *in vivo*, *i.e.*, capable of
10 replication under its own control.

A "vector" is a replicon, such as a plasmid, phage or cosmid, to which another DNA segment may be attached so as to bring about the replication of the attached segment.
15

A "cassette" refers to a segment of DNA that can be inserted into a vector at specific restriction sites. The segment of DNA encodes a polypeptide of interest, and the cassette and restriction sites are designed to ensure insertion of the cassette in the proper reading frame for transcription and translation.
20

"Heterologous" DNA refers to DNA not naturally located in the cell, or in a chromosomal site of the cell. Preferably, the heterologous DNA includes a gene foreign to the cell.

25 A cell has been "transfected" by exogenous or heterologous DNA when such DNA has been introduced inside the cell. A cell has been "transformed" by exogenous or heterologous DNA when the transfected DNA effects a phenotypic change. Preferably, the transforming DNA should be integrated (covalently linked) into chromosomal DNA making up the genome of the cell.

30

A "clone" is a population of cells derived from a single cell or common ancestor by mitosis.

- A "nucleic acid molecule" refers to the phosphate ester polymeric form of
- 5 ribonucleosides (adenosine, guanosine, uridine or cytidine; "RNA molecules") or deoxyribonucleosides (deoxyadenosine, deoxyguanosine, deoxythymidine, or deoxycytidine; "DNA molecules") in either single-stranded form, or a double-stranded helix. Double-stranded DNA-DNA, DNA-RNA and RNA-RNA helices are possible. The term nucleic acid molecule, and in particular DNA or RNA
- 10 molecule, refers only to the primary and secondary structure of the molecule, and does not limit it to any particular tertiary or quaternary forms. Thus, this term includes double-stranded DNA found, *inter alia*, in linear or circular DNA molecules (*e.g.*, restriction fragments), plasmids, and chromosomes. In discussing the structure of particular double-stranded DNA molecules, sequences may be
- 15 described herein according to the normal convention of giving only the sequence in the 5' to 3' direction along the nontranscribed strand of DNA (*i.e.*, the strand having a sequence homologous to the mRNA). A "recombinant DNA molecule" is a DNA molecule that has undergone a molecular biological manipulation.
- 20 A nucleic acid molecule is "hybridizable" to another nucleic acid molecule, such as a cDNA, genomic DNA, or RNA, when a single-stranded form of the nucleic acid molecule can anneal to the other nucleic acid molecule under the appropriate conditions of temperature and solution ionic strength (*see* Sambrook *et al.*, 1989, *supra*). The conditions of temperature and ionic strength determine the
- 25 "stringency" of the hybridization. For preliminary screening for homologous nucleic acids, low stringency hybridization conditions, corresponding to a T_m of 55°C, can be used, *e.g.*, 5x SSC, 0.1% SDS, 0.25% milk, and no formamide; or 30% formamide, 5x SSC, 0.5% SDS). Moderate stringency hybridization conditions correspond to a higher T_m , *e.g.*, 40% formamide, with 5x or 6x SCC.
- 30 High stringency hybridization conditions correspond to the highest T_m , *e.g.*, 50% formamide, 5x or 6x SCC. Hybridization requires that the two nucleic acids

contain complementary sequences, although depending on the stringency of the hybridization, mismatches between bases are possible. The appropriate stringency for hybridizing nucleic acids depends on the length of the nucleic acids and the degree of complementation, variables well known in the art. The greater the

5 degree of similarity or homology between two nucleotide sequences, the greater the value of T_m for hybrids of nucleic acids having those sequences. The relative stability (corresponding to higher T_m) of nucleic acid hybridizations decreases in the following order: RNA:RNA, DNA:RNA, DNA:DNA. For hybrids of greater than 100 nucleotides in length, equations for calculating T_m have been derived (*see*

10 Sambrook *et al.*, 1989, *supra*, 9.50-0.51). For hybridization with shorter nucleic acids, *i.e.*, oligonucleotides, the position of mismatches becomes more important, and the length of the oligonucleotide determines its specificity (*see* Sambrook *et al.*, 1989, *supra*, 11.7-11.8). Preferably a minimum length for a hybridizable nucleic acid is at least about 10 nucleotides; more preferably at least about 15

15 nucleotides; most preferably the length is at least about 20 nucleotides.

"Homologous recombination" refers to the insertion of a foreign DNA sequence of a vector in a chromosome. Preferably, the vector targets a specific chromosomal site for homologous recombination. For specific homologous recombination, the

20 vector will contain sufficiently long regions of homology to sequences of the chromosome to allow complementary binding and incorporation of the vector into the chromosome. Longer regions of homology, and greater degrees of sequence similarity, may increase the efficiency of homologous recombination.

25 A DNA "coding sequence" is a double-stranded DNA sequence which is transcribed and translated into a polypeptide in a cell *in vitro* or *in vivo* when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a start codon at the 5' (amino) terminus and a translation stop codon at the 3' (carboxyl) terminus. A coding sequence can

30 include, but is not limited to, prokaryotic sequences, cDNA from eukaryotic mRNA, genomic DNA sequences from eukaryotic (*e.g.*, mammalian) DNA, and

even synthetic DNA sequences. If the coding sequence is intended for expression in a eukaryotic cell, a polyadenylation signal and transcription termination sequence will usually be located 3' to the coding sequence.

5

Isolation of OB-R Coding and Flanking Sequences

The nucleic acids contemplated by the present invention include nucleic acids that code on expression for peptides such as those set forth in SEQ ID NOS:2, 4, 6, 8, and 10. Accordingly, while specific DNA has been isolated and sequenced in relation to the *OB-R* gene, any animal cell potentially can serve as the nucleic acid source for the molecular cloning of a gene encoding the polypeptides of the invention. The DNA may be obtained by standard procedures known in the art from cloned DNA (*e.g.*, a DNA "library"), by chemical synthesis, by cDNA cloning, or by the cloning of genomic DNA, or fragments thereof, purified from the desired cell (*See*, for example, Sambrook *et al.*, 1989, *supra*; Glover, 1985, *supra*). Clones derived from genomic DNA may contain regulatory and intronic DNA regions in addition to coding regions; clones derived from cDNA will not contain intron sequences. Whatever the source, the gene should be molecularly cloned into a suitable vector for propagation of the gene.

20

In the molecular cloning of the gene from genomic DNA, the genomic DNA can be amplified using primers selected from the cDNA sequences. Alternatively, DNA fragments are generated, some of which will encode the desired gene. The DNA may be cleaved at specific sites using various restriction enzymes. One may also use DNase in the presence of manganese to fragment the DNA, or the DNA can be physically sheared, as for example, by sonication. The linear DNA fragments can then be separated according to size by standard techniques, including but not limited to, agarose and polyacrylamide gel electrophoresis and column chromatography.

30

Once the DNA fragments are generated, identification of the specific DNA fragment containing the desired *OB-R*-gene may be accomplished in a number of ways. For example, if an amount of a portion of a *OB-R*-gene or its specific RNA, or a fragment thereof, is available and can be purified and labeled, the generated DNA fragments may be screened by nucleic acid hybridization to a labeled probe [Benton *et al.*, *Science*, **196**:180 (1977); Grunstein *et al.*, *Proc. Natl. Acad. Sci. USA*, **72**:3961 (1975)]. The present invention provides such nucleic acid probes, which can be conveniently prepared from the specific sequences disclosed herein, *e.g.*, a hybridizable probe having a nucleotide sequence corresponding to at least a 10, and preferably a 15, nucleotide fragment of the sequences depicted in SEQ ID NOS:1, 3, 5, 7, and 9. Preferably, a fragment is selected that is highly unique to the modulator peptides of the invention. Those DNA fragments with substantial homology to the probe will hybridize. As noted above, the greater the degree of homology, the more stringent the hybridization conditions that can be used. In one embodiment, low stringency hybridization conditions are used to identify a homologous modulator peptide. However, in a preferred aspect, and as demonstrated experimentally herein, a nucleic acid encoding a polypeptide of the invention will hybridize to a nucleic acid having a nucleotide sequence such as depicted in (SEQ ID NOS:1, 3, 5, 7, and 9), or a hybridizable fragment thereof, under moderately stringent conditions; more preferably, it will hybridize under high stringency conditions.

In another specific embodiment, the DNA of the invention can be identified using one of the PCR probes obtained by exon trapping and cDNA selection. For example, the primer pairs described in Example 3 can be used to will amplify a DNA of the invention.

Preferably, these primers will amplify DNA under moderately to high stringency conditions, *e.g.*, using pre-hybridization at 65° using Rapid-hyb buffer (Amersham Life Sciences), followed by hybridization for 6 hours at 65°, followed by washing first with 2XSSC/ 0.1% SDS for 30 min at room temperature (RT), and a second

wash at higher stringency with 0.3X SSC/ 0.1% SDS, RT, for 30 min. As will be appreciated by those of skill in the art, the stringency of the second wash is flexible and depends on the length of the probe and the degree of sequence similarity of each probe. For example, since human and mouse coding regions are
5 about 78% homologous, the same hybridization conditions may be employed with a lower the stringency second wash (*e.g.*, twice with 2XSSC/ 0.1%SDS, RT). If this results in no signal with no-background, hybridization can be attempted at a lower temperature (lower stringency), *e.g.*, 42°C. If there is too much background, the stringency of the second wash can be increased, (*e.g.* , 0.5 or
10 0.3X SSC, 0.1%SDS, RT). According to the invention, the above-noted PCR probes will define a nucleic acid molecule, *e.g.*, DNA, encoding OB-R from human as well as murine DNA libraries under similar hybridization conditions.

Alternatively, the presence of the gene may be detected by assays based on the
15 physical, chemical, or immunological properties of its expressed product. For example, cDNA clones, or DNA clones which hybrid-select the proper mRNAs, can be selected which produce a protein that, *e.g.*, has similar or identical electrophoretic migration, isoelectric focusing behavior, proteolytic digestion maps, tyrosine phosphatase activity, or antigenic properties as known for the
20 present OB-R. For example, antibodies of the instant invention can conveniently be used to screen for homologs of OB-R from other sources.

A gene encoding a polypeptide of the invention can also be identified by mRNA selection, *i.e.*, by nucleic acid hybridization followed by *in vitro* translation. In
25 this procedure, fragments are used to isolate complementary mRNAs by hybridization. Such DNA fragments may represent available, purified modulator DNA. Immunoprecipitation analysis or functional assays (*e.g.*, tyrosine phosphatase activity) of the *in vitro* translation products of the products of the isolated mRNAs identifies the mRNA and, therefore, the complementary DNA
30 fragments, that contain the desired sequences. In addition, specific mRNAs may

be selected by adsorption of polysomes isolated from cells to immobilized antibodies specifically directed against a modulator peptide.

A radiolabeled modulator peptide cDNA can be synthesized using the selected
5 mRNA (from the adsorbed polysomes) as a template. The radiolabeled mRNA or
cDNA may then be used as a probe to identify homologous modulator peptide
DNA fragments from among other genomic DNA fragments.

As mentioned above, a DNA sequence encoding weight modulator peptides as
10 disclosed herein can be prepared synthetically rather than cloned. The DNA
sequence can be designed with the appropriate codons for the OB-R amino acid
sequences. In general, one will select preferred codons for the intended host if the
sequence will be used for expression. The complete sequence may be assembled
from overlapping oligonucleotides prepared by standard methods and assembled
15 into a complete coding sequence. See, *e.g.*, Edge, *Nature*, **292**:756 (1981);
Nambair *et al.*, *Science*, **223**:1299 (1984); Jay *et al.*, *J. Biol. Chem.*, **259**:6311
(1984).

Synthetic DNA sequences allow convenient construction of genes which will
20 express OB-R analogs, as described above. Alternatively, DNA encoding analogs
can be made by site-directed mutagenesis of native *OB-R* genes or cDNAs, and
analogues can be made directly using conventional polypeptide synthesis.

A general method for site-specific incorporation of unnatural amino acids into
25 proteins is described in Noren *et al*, *Science*, **244**:182-188 (1989). This method
may be used to create analogs of the OB-R polypeptide with unnatural amino
acids.

Due to the degeneracy of nucleotide coding sequences, other DNA sequences
30 which encode substantially the same amino acid sequence as a *OB-R* gene may be
used in the practice of the present invention. These include but are not limited to

allelic genes, homologous genes from other species, and nucleotide sequences comprising all or portions of *OB-R* genes which are altered by the substitution of different codons that encode the same amino acid residue within the sequence, thus producing a silent change. Likewise, the OB-R derivatives of the invention
5 include, but are not limited to, those containing, as a primary amino acid sequence, all or part of the amino acid sequence of a OB-R protein including altered sequences in which functionally equivalent amino acid residues are substituted for residues within the sequence resulting in a conservative amino acid substitution, as described above in connection with OB-R analogs.

10

Non-coding Nucleic Acids

The present invention extends to the preparation of antisense nucleotides and ribozymes that may be used to interfere with the expression of the proteins at the translational level. This approach utilizes antisense nucleic acid and ribozymes to
15 block translation of a specific mRNA, either by masking that mRNA with an antisense nucleic acid or cleaving it with a ribozyme.

Antisense nucleic acids are DNA or RNA molecules that are complementary to at least a portion of a specific mRNA molecule [See Weintraub, *Sci. Am.*, **262**:40-46
20 (1990); Marcus-Sekura, *Anal. Biochem.*, **172**:289-295 (1988)]. In the cell, they hybridize to that mRNA, forming a double-stranded molecule. The cell does not translate an mRNA complexed in this double-stranded form. Therefore, antisense nucleic acids interfere with the expression of mRNA into protein. Oligomers of about fifteen nucleotides and molecules that hybridize to the AUG initiation codon
25 will be particularly efficient, since they are easy to synthesize and are likely to pose fewer problems than larger molecules when introducing them into weight modulator peptide-producing cells. Antisense methods have been used to inhibit the expression of many genes *in vitro* [(Marcus-Sekura, 1988 *supra*; Hambor *et al.*, *J. Exp. Med.*, **168**:1237-1245 (1988)].

30

Ribozymes are RNA molecules possessing the ability to specifically cleave other single-stranded RNA molecules in a manner somewhat analogous to DNA restriction endonucleases. Ribozymes were discovered from the observation that certain mRNAs have the ability to excise their own introns. By modifying the

5 nucleotide sequence of these RNAs, researchers have been able to engineer molecules that recognize specific nucleotide sequences in an RNA molecule and cleave it [Cech, *J. Am. Med. Assoc.*, **260**:3030-3034 (1988)]. Because ribozymes are sequence-specific, only mRNAs with particular sequences are inactivated.

10 Investigators have identified two types of ribozymes, *Tetrahymena*-type and "hammerhead"-type. *Tetrahymena*-type ribozymes recognize four-base sequences, while "hammerhead"-type recognize eleven- to eighteen-base sequences. The longer the recognition sequence, the more likely it is to occur exclusively in the target mRNA species. Therefore, hammerhead-type ribozymes are preferable to

15 *Tetrahymena*-type ribozymes for inactivating a specific mRNA species, and eighteen base recognition sequences are preferable to shorter recognition sequences.

The DNA sequences described herein may thus be used to prepare antisense

20 molecules against and ribozymes that cleave mRNAs for weight modulator proteins and their ligands, thus inhibiting expression of the *OB-R* gene, and leading to increased weight gain and adiposity.

In another embodiment, short oligonucleotides complementary to the coding and

25 complementary strands of the *OB-R* nucleic acid, or to non-coding regions of the *OB-R* gene 5', 3', or internal (intronic) to the coding region are provided by the present invention. Such nucleic acids are useful as probes, either as directly labeled oligonucleotide probes, or as primers for the polymerase chain reaction, for evaluating the presence of mutations in the *ob-r* gene, or the level of

30 expression of *OB-R* mRNA. Preferably, the non-coding nucleic acids of the invention are from the human *OB-R* gene.

In a specific embodiment, the non-coding nucleic acids provide for homologous recombination for integration of an amplifiable gene and/or other regulatory sequences in proximity to the *OB-R* gene, *e.g.*, to provide for higher levels of expression of the OB-R polypeptide, or to overcome a mutation in the *ob-r* gene regulatory sequences that prevent proper levels of expression of the OB-R polypeptide (See International Patent Publication WO 91/06666, published May 16, 1991 by Skoultchi; International Patent Publication No. WO 91/09955, published July 11, 1991 by Chappel; *see also* International Patent Publication No. WO 90/14092, published November 29, 1990, by Kucherlapati and Campbell).

10

Production of OB-R Polypeptide: Expression and Synthesis

Transcriptional and translational control sequences are DNA regulatory sequences, such as promoters, enhancers, terminators, and the like, that provide for the expression of a coding sequence in a host cell. In eukaryotic cells, polyadenylation signals are control sequences.

A coding sequence is "under the control" of transcriptional and translational control sequences in a cell when RNA polymerase transcribes the coding sequence into mRNA, which is then trans-RNA spliced and translated into the protein encoded by the coding sequence.

A "signal sequence" is included at the beginning of the coding sequence of a protein to be expressed on the surface of a cell. This sequence encodes a signal peptide, N-terminal to the mature polypeptide, that directs the host cell to translocate the polypeptide. The term "translocation signal sequence" is also used herein to refer to this sort of signal sequence. Translocation signal sequences can be found associated with a variety of proteins native to eukaryotes and prokaryotes, and are often functional in both types of organisms. According to the present invention, amino acid residues 1-27 of the murine and human OB-R polypeptides (*see* SEQ ID NOS:8, 10) comprise the signal peptide. In another

embodiment, amino acid residues 1-22 comprise the signal peptide [Tartaglia *et al.*, *Cell*, 83:1263 (1995)].

- A DNA sequence is "operatively linked" to an expression control sequence when the expression control sequence controls and regulates the transcription and translation of that DNA sequence. The term "operatively linked" includes having an appropriate start signal (*e.g.*, ATG) in front of the DNA sequence to be expressed and maintaining the correct reading frame to permit expression of the DNA sequence under the control of the expression control sequence and production of the desired product encoded by the DNA sequence. If a gene that one desires to insert into a recombinant DNA molecule does not contain an appropriate start signal, such a start signal can be inserted upstream (5') of and in reading frame with the gene.
- A "promoter sequence" is a DNA regulatory region capable of binding RNA polymerase in a cell and initiating transcription of a downstream (3' direction) coding sequence. For purposes of defining the present invention, the promoter sequence is bounded at its 3' terminus by the transcription initiation site and extends upstream (5' direction) to include the minimum number of bases or elements necessary to initiate transcription at levels detectable above background. Within the promoter sequence will be found a transcription initiation site (conveniently defined for example, by mapping with nuclease S1), as well as protein binding domains (consensus sequences) responsible for the binding of RNA polymerase.
- Another feature of this invention is the expression of the DNA sequences disclosed herein. As is well known in the art, DNA sequences may be expressed by operatively linking them to an expression control sequence in an appropriate expression vector and employing that expression vector to transform an appropriate unicellular host.

Such operative linking of a DNA sequence of this invention to an expression control sequence, of course, includes, if not already part of the DNA sequence, the provision of an initiation codon, ATG, in the correct reading frame upstream of the DNA sequence.

5

- A wide variety of host/expression vector combinations may be employed in expressing the DNA sequences of this invention. Useful expression vectors, for example, may consist of segments of chromosomal, non-chromosomal, and synthetic DNA sequences. Suitable vectors include derivatives of SV40 and
- 10 known bacterial plasmids, *e.g.*, *E. coli* plasmids col El, pCR1, pBR322, pMB9, pUC or pUC plasmid derivatives, *e.g.*, pGEX vectors, pET vectors, pmal-c, pFLAG, *etc.*, and their derivatives, plasmids such as RP4; phage DNAs, *e.g.*, the numerous derivatives of phage λ , such as NM989, and other phage DNA, *e.g.*, M13 and filamentous single-stranded phage DNA; yeast plasmids such as the 2 μ
- 15 plasmid or derivatives thereof; vectors useful in eukaryotic cells, such as vectors useful in insect or mammalian cells; vectors derived from combinations of plasmids and phage DNAs, such as plasmids that have been modified to employ phage DNA or other expression control sequences; and the like.
- 20 Any of a wide variety of expression control sequences -- sequences that control the expression of a DNA sequence operatively linked to it -- may be used in these vectors to express the DNA sequences of this invention. Such useful expression control sequences include, for example, the early or late promoters of SV40, CMV, vaccinia, polyoma or adenovirus, the *lac* system, the *trp* system, the *TAC*
- 25 system, the *TRC* system, the *LTR* system, the major operator and promoter regions of phage λ , the control regions of fd coat protein, the promoter for 3-phosphoglycerate kinase or other glycolytic enzymes, the promoters of acid phosphatase (*e.g.*, Pho5), the AOX 1 promoter of methylotrophic yeast, the promoters of the yeast α -mating factors, and other sequences known to control the
- 30 expression of genes of prokaryotic or eukaryotic cells or their viruses, and various combinations thereof.

A wide variety of unicellular host cells are also useful in expressing the DNA sequences of this invention. These hosts may include well known eukaryotic and prokaryotic hosts, such as strains of *E. coli*, *Pseudomonas*, *Bacillus*, *Streptomyces*; fungi such as yeasts (*Saccharomyces*, and methylotrophic yeast such as *Pichia*,
5 *Candida*, *Hansenula*, and *Torulopsis*); and animal cells, such as CHO, R1.1, B-W and LM cells, African Green Monkey kidney cells (*e.g.*, COS 1, COS 7, BSC1, BSC40, and BMT10), insect cells (*e.g.*, Sf9), and human cells and plant cells in tissue culture.

- 10 It will be understood that not all vectors, expression control sequences and hosts will function equally well to express the DNA sequences of this invention. Neither will all hosts function equally well with the same expression system. However, one skilled in the art will be able to select the proper vectors, expression control sequences, and hosts without undue experimentation to
15 accomplish the desired expression without departing from the scope of this invention. For example, in selecting a vector, the host must be considered because the vector must function in it. The vector's copy number, the ability to control that copy number, and the expression of any other proteins encoded by the vector, such as antibiotic markers, will also be considered.

20

- In selecting an expression control sequence, a variety of factors will normally be considered. These include, for example, the relative strength of the system, its controllability, and its compatibility with the particular DNA sequence or gene to be expressed, particularly as regards potential secondary structures. Suitable
25 unicellular hosts will be selected by consideration of, *e.g.*, their compatibility with the chosen vector, their secretion characteristics, their ability to fold proteins correctly, and their fermentation requirements, as well as the toxicity to the host of the product encoded by the DNA sequences to be expressed, and the ease of purification of the expression products.

30

Considering these and other factors, a person skilled in the art will be able to construct a variety of vector/expression control sequence/host combinations that will express the DNA sequences of this invention on fermentation or in large scale animal culture.

5

In a specific embodiment, an OB-R fusion protein can be expressed. An OB-R fusion protein comprises at least a functionally active portion of a non-OB-R protein joined via a peptide bond to at least a functionally active portion of an OB polypeptide. The non-OB-R sequences can be amino- or carboxy-terminal to the
10 OB-R sequences. For example, in preparing "artificial" receptors, joining the OB-R encoding coding domain for the leptin binding (extracytoplasmic) portion at the 5' position will yield a protein that binds leptin and mediates some other action based on the non-OB-R protein's activity. Conversely, joining a different protein (such as a growth factor, cytokine, or hormone receptor binding coding domain)
15 5' to a OB-R cytoplasmic coding domain (containing "Box 1" and "Box 2") will allow for activation via OB-R upon binding a different ligand than leptin. In another embodiment, a chimeric construct may simply facilitate expression of OB-R.

20 In another aspect, the pGEX vector [Smith *et al.*, Gene 67:31-40 (1988)] can be used. This vector fuses the schistosoma japonicum glutathione S-transferase cDNA to the sequence of interest. Bacterial proteins are harvested and recombinant proteins can be quickly purified on a reduced glutathione affinity column. The GST carrier can subsequently be cleaved from fusion proteins by
25 digestion with site-specific proteases. After cleavage, the carrier and uncleaved fusion protein can be removed by absorption on glutathione agarose. Difficulty with the system occasionally arises when the encoded protein is insoluble in aqueous solutions.

30 Expression of recombinant proteins in bacterial systems may result in incorrect folding of the expressed protein, requiring refolding. The recombinant protein can

be refolded prior to or after cleavage to form a functionally active OB polypeptide. The OB polypeptide may be refolded by the steps of (i) incubating the protein in a denaturing buffer that contains a reducing agent, and then (ii) incubating the protein in a buffer that contains an oxidizing agent, and preferably also contains a protein stabilizing agent or a chaotropic agent, or both. Suitable redox (reducing/oxidizing) agent pairs include, but are not limited to, reduced glutathione/glutathione disulfide, cystine/cysteine, cystamine/cysteamine, and 2-mercaptoethanol/2-hydroxyethyl disulfide. In a particular aspect, the fusion protein can be solubilized in a denaturant, such as urea, prior to exchange into the reducing buffer. In preferred embodiment, the protein is also purified, *e.g.*, by ion exchange or Ni-chelation chromatography, prior to exchange into the reducing buffer. Denaturing agents include but are not limited to urea and guanidine-HCl. The recombinant protein is then diluted about at least 10-fold, more preferably about 100-fold, into an oxidizing buffer that contains an oxidizing agent, such as but not limited to 0.1 M Tris-HCl, pH 8.0, 1 mM EDTA, 0.15 M NaCl, 0.3 M oxidized glutathione. The fusion protein is then incubated for about 1 to about 24 hours, preferably about 2 to about 16 hours, at room temperature in the oxidizing buffer. The oxidizing buffer may comprise a protein stabilizing agent, *e.g.*, a sugar, an alcohol, or ammonium sulfate. The oxidizing buffer may further comprises a chaotropic agent at low concentration, to destabilize incorrect intermolecular interactions and thus promote proper folding. Suitable chaotropic agents include but are not limited to a detergent, a polyol, L-arginine, guanidine-HCl and polyethylene glycol (PEG). It is important to use a low enough concentration of the chaotropic agent to avoid denaturing the protein. The refolded protein can be concentrated by at least about 10-fold, more preferably by the amount it was diluted into the oxidizing buffer.

Alternatively, the invention contemplates periplasmic expression of a protein of the invention.

Bacterial fermentation processes can also result in a recombinant protein preparation that contains unacceptable levels of endotoxins. Therefore, the invention contemplates removal of such endotoxins, *e.g.*, by using endotoxin-specific antibodies or other endotoxin binding molecules. The presence of
 5 endotoxins can be determined by standard techniques, such as by employing E-TOXATE Reagents (Sigma, St. Louis, Missouri), or with bioassays.

In addition to the specific example, the present inventors contemplate use of baculovirus, mammalian, and yeast expression systems to express the ob protein.
 10 For example, in baculovirus expression systems, both non-fusion transfer vectors, such as but not limited to pVL941 (*Bam*H1 cloning site; Summers), pVL1393 (*Bam*H1, *Sma*I, *Xba*I, *Eco*R1, *Not*I, *Xma*III, *Bgl*II, and *Pst*I cloning site; Invitrogen), pVL1392 (*Bgl*II, *Pst*I, *Not*I, *Xma*III, *Eco*RI, *Xba*I, *Sma*I, and *Bam*H1 cloning site; Summers and Invitrogen), and pBlueBacIII (*Bam*H1, *Bgl*II, *Pst*I,
 15 *Nco*I, and *Hind*III cloning site, with blue/white recombinant screening possible; Invitrogen), and fusion transfer vectors, such as but not limited to pAc700 (*Bam*H1 and *Kpn*I cloning site, in which the *Bam*H1 recognition site begins with the initiation codon; Summers), pAc701 and pAc702 (same as pAc700, with different reading frames), pAc360 (*Bam*H1 cloning site 36 base pairs downstream
 20 of a polyhedrin initiation codon; Invitrogen(195)), and pBlueBacHisA, B, C (three different reading frames, with *Bam*H1, *Bgl*II, *Pst*I, *Nco*I, and *Hind*III cloning site, an N-terminal peptide for ProBond purification, and blue/white recombinant screening of plaques; Invitrogen (220)).

25 Mammalian expression vectors contemplated for use in the invention include vectors with inducible promoters, such as the dihydrofolate reductase (DHFR) promoter, *e.g.*, any expression vector with a *DHFR* expression vector, or a *DHFR*/methotrexate co-amplification vector, such as pED [*Pst*I, *Sal*I, *Sba*I, *Sma*I, and *Eco*RI cloning site, with the vector expressing both the cloned gene and
 30 *DHFR*; see Kaufman, *Current Protocols in Molecular Biology*, 16.12 (1991)]. Alternatively, a glutamine synthetase/methionine sulfoximine co-amplification

vector, such as pEE14 (*HindIII*, *XbaI*, *SmaI*, *SbaI*, *EcoRI*, and *BclI* cloning site, in which the vector expresses glutamine synthase and the cloned gene; Celltech). In another embodiment, a vector that directs episomal expression under control of Epstein Barr Virus (EBV) can be used, such as pREP4 (*BamHI*, *SfiI*, *XhoI*, *NotI*,
5 *NheI*, *HindIII*, *NheI*, *PvuII*, and *KpnI* cloning site, constitutive RSV-LTR promoter, hygromycin selectable marker; Invitrogen), pCEP4 (*BamHI*, *SfiI*, *XhoI*, *NotI*, *NheI*, *HindIII*, *NheI*, *PvuII*, and *KpnI* cloning site, constitutive hCMV immediate early gene, hygromycin selectable marker; Invitrogen), pMEP4 (*KpnI*, *PvuI*, *NheI*, *HindIII*, *NotI*, *XhoI*, *SfiI*, *BamHI* cloning site, inducible
10 methallothionein IIa gene promoter, hygromycin selectable marker: Invitrogen), pREP8 (*BamHI*, *XhoI*, *NotI*, *HindIII*, *NheI*, and *KpnI* cloning site, RSV-LTR promoter, histidinol selectable marker; Invitrogen), pREP9 (*KpnI*, *NheI*, *HindIII*, *NotI*, *XhoI*, *SfiI*, and *BamHI* cloning site, RSV-LTR promoter, G418 selectable marker; Invitrogen), and pEBVHis (RSV-LTR promoter, hygromycin selectable
15 marker, N-terminal peptide purifiable via ProBond resin and cleaved by enterokinase; Invitrogen). Selectable mammalian expression vectors for use in the invention include pRc/CMV (*HindIII*, *BstXI*, *NotI*, *SbaI*, and *ApaI* cloning site, G418 selection; Invitrogen), pRc/RSV (*HindIII*, *SpeI*, *BstXI*, *NotI*, *XbaI* cloning site, G418 selection; Invitrogen), and others. Vaccinia virus mammalian
20 expression vectors (*see*, Kaufman, 1991, *supra*) for use according to the invention include but are not limited to pSC11 (*SmaI* cloning site, TK- and β -gal selection), pMJ601 (*SalI*, *SmaI*, *AflI*, *NarI*, *BspMII*, *BamHI*, *ApaI*, *NheI*, *SacII*, *KpnI*, and *HindIII* cloning site; TK- and β -gal selection), and pTKgptF1S (*EcoRI*, *PstI*, *SalI*, *AccI*, *HindII*, *SbaI*, *BamHI*, and *HpaI* cloning site, TK or XPRT selection).
25

Yeast expression systems can also be used according to the invention to express OB polypeptide. For example, the non-fusion pYES2 vector (*XbaI*, *SphI*, *ShoI*, *NotI*, *GstXI*, *EcoRI*, *BstXI*, *BamHI*, *SacI*, *KpnI*, and *HindIII* cloning sit;
Invitrogen) or the fusion pYESHisA, B, C (*XbaI*, *SphI*, *ShoI*, *NotI*, *BstXI*, *EcoRI*,
30 *BamHI*, *SacI*, *KpnI*, and *HindIII* cloning site, N-terminal peptide purified with

ProBond resin and cleaved with enterokinase; Invitrogen), to mention just two, can be employed according to the invention.

5 It is further intended that body weight modulator polypeptides and analogs may be prepared from nucleotide sequences derived within the scope of the present invention.

In addition to recombinant expression of OB-R polypeptide, the present invention envisions and fully enables preparation of OB-R polypeptide, or fragments thereof, 10 using the well known and highly developed techniques of solid phase peptide synthesis. The invention contemplates using both the popular Boc and Fmoc, as well as other protecting group strategies, for preparing ob polypeptide or fragments thereof. Various techniques for refolding and oxidizing the cysteine side chains to form a disulfide bond are also well-known in the art.

15

Antibodies to the OB-R Polypeptide

According to the invention, OB-R polypeptide produced recombinantly or by chemical synthesis, and fragments or other derivatives or analogs thereof, 20 including fusion proteins, may be used as an immunogen to generate antibodies that recognize the OB-R polypeptide. Such antibodies include but are not limited to polyclonal, monoclonal, chimeric, single chain, Fab fragments, and an Fab expression library.

25 A molecule is "antigenic" when it is capable of specifically interacting with an antigen recognition molecule of the immune system, such as an immunoglobulin (antibody) or T cell antigen receptor. An antigenic polypeptide contains at least about 5, and preferably at least about 10, amino acids. An antigenic portion of a molecule can be that portion that is immunodominant for antibody or T cell 30 receptor recognition, or it can be a portion used to generate an antibody to the molecule by conjugating the antigenic portion to a carrier molecule for

immunization. A molecule that is antigenic need not be itself immunogenic, *i.e.*, capable of eliciting an immune response without a carrier.

An "antibody" is any immunoglobulin, including antibodies and fragments thereof,
5 that binds a specific epitope. The term encompasses polyclonal, monoclonal, and
chimeric antibodies, the last mentioned described in further detail in U.S. Patent
Nos. 4,816,397 and 4,816,567, as well as antigen binding portions of antibodies,
including Fab, F(ab')₂ and F(v) (including single chain antibodies). Accordingly,
the phrase "antibody molecule" in its various grammatical forms as used herein
10 contemplates both an intact immunoglobulin molecule and an immunologically
active portion of an immunoglobulin molecule containing the antibody combining
site. An "antibody combining site" is that structural portion of an antibody
molecule comprised of heavy and light chain variable and hypervariable regions
that specifically binds antigen.

15 Exemplary antibody molecules are intact immunoglobulin molecules, substantially
intact immunoglobulin molecules and those portions of an immunoglobulin
molecule that contains the paratope, including those portions known in the art as
Fab, Fab', F(ab')₂ and F(v), which portions are preferred for use in the
20 therapeutic methods described herein.

Fab and F(ab')₂ portions of antibody molecules are prepared by the proteolytic
reaction of papain and pepsin, respectively, on substantially intact antibody
molecules by methods that are well-known. See for example, U.S. Patent No.
25 4,342,566 to Theofilopolous *et al.* Fab' antibody molecule portions are also well-
known and are produced from F(ab')₂ portions followed by reduction of the
disulfide bonds linking the two heavy chain portions as with mercaptoethanol, and
followed by alkylation of the resulting protein mercaptan with a reagent such as
iodoacetamide. An antibody containing intact antibody molecules is preferred
30 herein.

The phrase "monoclonal antibody" in its various grammatical forms refers to an antibody having only one species of antibody combining site capable of immunoreacting with a particular antigen. A monoclonal antibody thus typically displays a single binding affinity for any antigen with which it immunoreacts. A
5 monoclonal antibody may therefore contain an antibody molecule having a plurality of antibody combining sites, each immunospecific for a different antigen; *e.g.*, a bispecific (chimeric) monoclonal antibody.

The term "adjuvant" refers to a compound or mixture that enhances the immune
10 response to an antigen. An adjuvant can serve as a tissue depot that slowly releases the antigen and also as a lymphoid system activator that non-specifically enhances the immune response [Hood *et al.*, in *Immunology*, p. 384, Second Ed., Benjamin/Cummings, Menlo Park, California (1984)]. Often, a primary challenge with an antigen alone, in the absence of an adjuvant, will fail to elicit a humoral
15 or cellular immune response. Adjuvants include, but are not limited to, complete Freund's adjuvant, incomplete Freund's adjuvant, saponin, mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil or hydrocarbon emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially useful human adjuvants such as BCG
20 (*bacille Calmette-Guerin*) and *Corynebacterium parvum*. Preferably, the adjuvant is pharmaceutically acceptable.

Various procedures known in the art may be used for the production of polyclonal antibodies to OB-R polypeptide, or fragment, derivative or analog thereof. For
25 the production of antibody, various host animals can be immunized by injection with the OB-R polypeptide, or a derivative (*e.g.*, fragment or fusion protein) thereof, including but not limited to rabbits, mice, rats, sheep, goats, etc. In one embodiment, the OB-R polypeptide or fragment thereof can be conjugated to an immunogenic carrier, *e.g.*, bovine serum albumin (BSA) or keyhole limpet
30 hemocyanin (KLH). Specific OB-R antigenic fragments are disclosed in Example 2, *infra*. Various adjuvants may be used to increase the immunological response,

depending on the host species, including but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially useful human adjuvants such as

5 BCG (*bacille Calmette-Guerin*) and *Corynebacterium parvum*.

For preparation of monoclonal antibodies directed toward the OB-R polypeptide, or fragment, analog, or derivative thereof, any technique that provides for the production of antibody molecules by continuous cell lines in culture may be used.

10 These include but are not limited to the hybridoma technique originally developed by Kohler *et al.*, *Nature*, **256**:495-497 (1975), as well as the trioma technique, the human B-cell hybridoma technique [Kozbor *et al.*, *Immunology Today*, **4**:72 (1983)], and the EBV-hybridoma technique to produce human monoclonal antibodies [Cole *et al.*, in *Monoclonal Antibodies and Cancer Therapy*, pp. 77-96,

15 Alan R. Liss, Inc., (1985)]. Immortal, antibody-producing cell lines can be created by techniques other than fusion, such as direct transformation of B lymphocytes with oncogenic DNA, or transfection with Epstein-Barr virus. *See, e.g.*, M. Schreier *et al.*, "Hybridoma Techniques" (1980); Hammerling *et al.*, "Monoclonal Antibodies And T-cell Hybridomas" (1981); Kennett *et al.*,

20 "Monoclonal Antibodies" (1980); *see also* U.S. Patent Nos. 4,341,761; 4,399,121; 4,427,783; 4,444,887; 4,451,570; 4,466,917; 4,472,500; 4,491,632; and 4,493,890.

In an additional embodiment of the invention, monoclonal antibodies can be

25 produced in germ-free animals utilizing recent technology (PCT/US90/02545). According to the invention, human antibodies may be used and can be obtained by using human hybridomas [Cote *et al.*, *Proc. Natl. Acad. Sci. USA*, **80**:2026-2030 (1983)] or by transforming human B cells with EBV virus *in vitro* (Cole *et al.*, 1985, *supra*). In fact, according to the invention, techniques developed for the

30 production of "chimeric antibodies" [Morrison *et al.*, *J. Bacteriol.*, 159-870 (1984); Neuberger *et al.*, *Nature*, **312**:604-608 (1984); Takeda *et al.*, *Nature*,

314:452-454 (1985)] by splicing the genes from a mouse antibody molecule specific for an ob polypeptide together with genes from a human antibody molecule of appropriate biological activity can be used; such antibodies are within the scope of this invention. Such human or humanized chimeric antibodies are
5 preferred for use in therapy of human diseases or disorders (described *infra*), since the human or humanized antibodies are much less likely than xenogenic antibodies to induce an immune response, in particular an allergic response, themselves.

According to the invention, techniques described for the production of single chain
10 antibodies (U.S. Patent 4,946,778) can be adapted to produce OB polypeptide-specific single chain antibodies. An additional embodiment of the invention utilizes the techniques described for the construction of Fab expression libraries [Huse *et al.*, *Science*, 246:1275-1281 (1989)] to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity for an ob polypeptide, or
15 its derivatives, or analogs.

Antibody fragments which contain the idiotype of the antibody molecule can be generated by known techniques. For example, such fragments include but are not limited to: the F(ab')₂ fragment which can be produced by pepsin digestion of the
20 antibody molecule; the Fab' fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragment, and the Fab fragments which can be generated by treating the antibody molecule with papain and a reducing agent.

In the production of antibodies, screening for the desired antibody can be
25 accomplished by techniques known in the art, *e.g.*, radioimmunoassay, ELISA (enzyme-linked immunosorbent assay), "sandwich" immunoassays, immunoradiometric assays, gel diffusion precipitin reactions, immunodiffusion assays, *in situ* immunoassays (using colloidal gold, enzyme or radioisotope labels, for example), Western blots, precipitation reactions, agglutination assays (*e.g.*, gel
30 agglutination assays, hemagglutination assays), complement fixation assays, immunofluorescence assays, protein A assays, and immunoelectrophoresis assays,

etc. In one embodiment, antibody binding is detected by detecting a label on the primary antibody. In another embodiment, the primary antibody is detected by detecting binding of a secondary antibody or reagent to the primary antibody. In a further embodiment, the secondary antibody is labeled. Many means are known in the art for detecting binding in an immunoassay and are within the scope of the present invention. For example, to select antibodies which recognize a specific epitope of an OB polypeptide, one may assay generated hybridomas for a product which binds to an OB polypeptide fragment containing such epitope. For selection of an antibody specific to an OB polypeptide from a particular species of animal, one can select on the basis of positive binding with OB polypeptide expressed by or isolated from cells of that species of animal.

The foregoing antibodies can be used in methods known in the art relating to the localization and activity of the OB-R polypeptide, *e.g.*, for Western blotting, imaging OB-R polypeptide *in situ*, measuring levels thereof in appropriate physiological samples, etc.

In a specific embodiment, antibodies that agonize or antagonize the activity of OB-R polypeptide can be generated. Such antibodies can be tested using the assays described *infra* for identifying ligands.

In a particular aspect, antibodies are developed by immunizing rabbits with synthetic peptides predicted by the protein sequence or with recombinant proteins made using bacterial expression vectors. The choice of synthetic peptides is made after careful analysis of the predicted protein structure, as described above. In particular, peptide sequences between putative cleavage sites are chosen. Synthetic peptides are conjugated to a carrier such as KLH hemocyanin or BSA using carbodiimide and used in Freund's adjuvant to immunize rabbits. In order to prepare recombinant protein, the pGEX vector can be used to express the polypeptide (Smith *et al.*, 1988, *supra*). Alternatively, one can use only

hydrophilic domains to generate the fusion protein. The expressed protein will be prepared in quantity and used to immunize rabbits in Freund's adjuvant.

- In a specific embodiment, *infra*, peptides corresponding to amino acid residues
- 5 145-158, 465-484, and 863-881 (from the murine OB-R polypeptide depicted in any one of SEQ ID NOS:8, 10) can be generated by solid phase peptide synthesis, conjugated to a carrier such as KLH, and used to immunize rabbits, rats, goats, chickens, etc.
- 10 In another specific embodiment, recombinant OB-R polypeptide is used to immunize chickens, and the chicken anti-OB-R antibodies are recovered from egg yolk, *e.g.*, by affinity purification on an OB-R-column. Preferably, chickens used in immunization are kept under specific pathogen free (SPF) conditions.
- 15 In yet another embodiment, recombinant OB-R polypeptide is used to immunize rabbits, and the polyclonal antibodies are immunopurified prior to further use. The purified antibodies are particularly useful for semi-quantitative assays, particularly for detecting the presence of the circulating (soluble) splice form(s) of OB-R polypeptide in serum or plasma.
- 20 Panels of monoclonal antibodies produced against modulator peptides can be screened for various properties; *i.e.*, isotype, epitope, affinity, etc. Of particular interest are monoclonal antibodies that neutralize the activity of the modulator peptides. Such monoclonals can be readily identified in activity assays for the
- 25 weight modulators. High affinity antibodies are also useful when immunoaffinity purification of native or recombinant polypeptide is desired.

Preferably, the anti-modulator antibody used in the diagnostic and therapeutic methods of this invention is an affinity-purified polyclonal antibody. More

30 preferably, the antibody is a monoclonal antibody (mAb). In addition, it is

preferable for the anti-modulator antibody molecules used herein be in the form of Fab, Fab', F(ab')₂ or F(v) portions of whole antibody molecules.

Diagnostics

5

The present invention also relates to a variety of diagnostic applications, including methods for detecting the presence of conditions and/or stimuli that impact upon abnormalities in body weight or adiposity, by reference to their ability to elicit the activities which are mediated by the present OB-R polypeptides. As mentioned
10 earlier, the peptides can be used to produce antibodies to themselves by a variety of known techniques, and such antibodies could then be isolated and utilized as in tests for the presence of particular transcriptional activity in suspect target cells. Alternatively, the nucleic acids of the invention can be employed in diagnosis.

15

Antibody-based Diagnostics

As suggested earlier, a diagnostic method useful in the present invention comprises examining a cellular sample or medium by means of an assay including an effective amount of an OB-R binding partner, such as an anti-modulator antibody or leptin, preferably an affinity-purified polyclonal antibody, and more preferably
20 a mAb. In addition, it is preferable for the antibody molecules used herein be in the form of Fab, Fab', F(ab')₂ or F(v) portions or whole antibody molecules. As previously discussed, patients capable of benefiting from this method include those suffering from cancer, AIDS, obesity or other conditions where abnormal body weight is an element of the condition.

25

Also, antibodies including both polyclonal and monoclonal antibodies, may possess certain diagnostic applications and may for example, be utilized for the purpose of detecting and/or measuring conditions where abnormalities in body weight are or may be likely to develop.

30

The diagnostic methods can be used to detect OB-R in a biological sample from an individual. The biological sample can be a biological fluid, such as but not limited to, blood, serum, plasma, interstitial fluid, plural effusions, urine, cerebrospinal fluid, and the like. Preferably, soluble OB-R is detected in serum or urine, which
5 are both readily obtained. Alternatively, OB-R can be detected from cellular sources, such as, but not limited to, brain tissue biopsies, adipocytes, testes, heart, and the like. For example, cells can be obtained from an individual by biopsy and lysed, *e.g.*, by freeze-thaw cycling, or treatment with a mild cytolytic detergent such as, but not limited to, TRITON X-100®, digitonin, NONIDET P
10 (NP)-40®, saponin, and the like, or combinations thereof (*see, e.g.*, International Patent Publication WO 92/08981, published May 29, 1992). In yet another embodiment, samples containing both cells and body fluids can be used (*see ibid.*).

The presence of OB-R in cells or in a biological fluid can be ascertained by the
15 usual immunological procedures applicable to such determinations. A number of useful procedures are known. Three such procedures which are especially useful utilize either the OB-R (particularly the secreted splice form) labeled with a detectable label, antibody Ab₁ labeled with a detectable label, or antibody Ab₂ labeled with a detectable label.

20

The procedures and their application are all familiar to those skilled in the art and accordingly may be utilized within the scope of the present invention. For example, a "competitive" procedure, is described in U.S. Patent Nos. 3,654,090 and 3,850,752. A "sandwich" procedure, is described in U.S. Patent Nos. RE
25 31,006 and 4,016,043. Still other procedures are known such as the "double antibody", or "DASP" procedure.

The labels most commonly employed for these studies are radioactive elements, enzymes, chemicals which fluoresce when exposed to ultraviolet light, and others.
30

A number of fluorescent materials are known and can be utilized as labels. These include, for example, fluorescein, rhodamine and auramine. A particular detecting material is anti-rabbit antibody prepared in goats and conjugated with fluorescein through an isothiocyanate.

5

The radioactive label can be detected by any of the currently available counting procedures. The preferred isotope may be selected from ^3H , ^{14}C , ^{32}P , ^{35}S , ^{36}Cl , ^{51}Cr , ^{57}Co , ^{58}Co , ^{59}Fe , ^{90}Y , ^{125}I , ^{131}I , and ^{186}Re .

- 10 Enzyme labels are likewise useful, and can be detected by any of the presently utilized colorimetric, spectrophotometric, fluorospectrophotometric, amperometric or gasometric techniques. The enzyme is conjugated by reaction with bridging molecules such as carbodiimides, diisocyanates, glutaraldehyde and the like. Many enzymes which can be used in these procedures are known and can be
- 15 utilized. The preferred are peroxidase, β -glucuronidase, β -D-glucosidase, β -D-galactosidase, urease, glucose oxidase plus peroxidase and alkaline phosphatase. U.S. Patent Nos. 3,654,090; 3,850,752; and 4,016,043 are referred to by way of example for their disclosure of alternate labeling material and methods.

20

- In a further embodiment of this invention, test kits suitable for use by a medical specialist may be prepared to determine the presence or absence of OB-R in suspected target cells or biological fluids. In accordance with the testing techniques discussed above, one class of such kits will contain at least the labeled
- 25 OB-R polypeptide or its binding partner, for instance an antibody specific thereto, and directions, of course, depending upon the method selected, e.g., "competitive," "sandwich," "DASP" and the like. The kits may also contain peripheral reagents such as buffers, stabilizers, etc.

Nucleic Acid-based Diagnostics

As demonstrated in the examples, *infra*, nucleic acids of the invention can be used to detect defects associated with defects in the OB-R polypeptide associated with an obese phenotypes. For example, nucleic acid probes (*e.g.*, in Northern analysis or RT-PCR analysis) can be used to determine whether an obese phenotype is associated with lack of expression of *OB-R* mRNA, or expression of non-functional *OB-R* mRNA, *e.g.*, as in *db/db* mice (where the deficiency results from lack of an effective leptin receptor), or where a mutation yields a non-transcribed mRNA. Moreover, the nucleic acid-based diagnostic techniques of the invention can be used in conjunction with antibody-based techniques to further develop a molecular understanding of obese or anorexic phenotypes.

Human cDNA clones may be sequenced. This facilitates the determination of the complete sequence of the human gene. DNA sequences from the introns of the human *OB-R* gene may thus be obtained, and these can be used to prepare PCR primers to PCR amplify the coding sequence of the *OB-R* gene from human genomic DNA so as to identify mutations or allelic variants of the *OB-R* gene, all in accordance with protocols described in detail earlier herein.

The current hypothesis is that heterozygous mutations in the *DB* gene will be associated with mild/moderate obesity while homozygous mutations would be associated with severe obesity. This would allow the ascertainment of people at risk for the development of obesity and make possible the application of drug treatment and/or lifestyle changes before an increased body weight is fully developed.

Alternatively, the presence of microsatellites that segregate with mutant forms of human *ob-r* can be used for diagnosis. Various PCR primers, can be used in this respect.

The *OB-R* gene may also be useful diagnostically for measurements of its encoded RNA and protein in nutritional disorders. It will be of importance to know, in a particular nutritional disorder, whether *OB-R* RNA and/or its encoded protein is upregulated or downregulated. Thus, if an obese person has increased levels of

5 *OB-R*, it would appear that the problem is downstream of *OB-R*, while if *OB-R* expression is reduced, it would appear that inappropriately low levels of *OB* may be cause of obesity (whether or not the defect is in the *OB-R* gene). Conversely, if a cancer or AIDS patient who lost weight had elevated levels of *OB-R*, it may be concluded that inappropriately high expression of *OB-R* is responsible for the

10 weight loss.

The present invention is concerned with not only inappropriate levels of expression of *OB-R*, but also with expression of non-functional or dysfunctional splice forms. The nucleic acid diagnostics of the invention provide for determining whether the

15 predominantly expressed form is dysfunctional, *e.g.*, for signal transduction. As demonstrated in the Examples, *infra*, *db* mutant mice (C57BL/Ks *db/db*) express a longer *OB-R* mRNA (as determined by RT-PCR).

Therapeutics

20 The polypeptides, nucleic acids, and antibodies of the invention have significant therapeutic potential. Preferably, a therapeutically effective amount of such an agent (*e.g.*, soluble form of the protein, or DNA for gene therapy, or an antisense nucleic acid for antagonizing leptin activity) is administered in a pharmaceutically

25 acceptable carrier, diluent, or excipient.

The phrase "pharmaceutically acceptable" refers to molecular entities and compositions that are physiologically tolerable and do not typically produce an allergic or similarly untoward reaction, such as gastric upset, dizziness and the

30 like, when administered to a human. In one embodiment, as used herein, the term "pharmaceutically acceptable" may mean approved by a regulatory agency of the

federal or a state government or listed in the *U.S. Pharmacopeia* or other generally recognized pharmacopeia for use in animals, and more particularly in humans. The term "carrier" refers to a diluent, adjuvant, excipient, or vehicle with which the compound is administered. Such pharmaceutical carriers can be
5 sterile liquids, such as water and oils, including those of petroleum, animal, vegetable or synthetic origin, such as peanut oil, soybean oil, mineral oil, sesame oil and the like. Water or solution saline solutions and aqueous dextrose and glycerol solutions are preferably employed as carriers, particularly for injectable solutions. Suitable pharmaceutical carriers are described in Martin, *Remington's*
10 *Pharmaceutical Sciences*, 18th Ed., Mack Publishing Co., Easton, PA, (1990).

The phrase "therapeutically effective amount" is used herein to mean an amount sufficient to reduce by at least about 15%, preferably by at least 50%, more preferably by at least 90%, and most preferably prevent, a clinically significant
15 deficit in the activity, function and response of the host. Alternatively, a therapeutically effective amount is sufficient to cause an improvement in a clinically significant condition in the host. Modulation of OB-R activity can be useful for reducing body weight (by increasing its activity) or increasing body weight (by decreasing its activity).

20 Administration of recombinant soluble OB-R polypeptide corresponding to OB-Re is expected to result in weight loss, in particular, a decrease in fat tissue. Soluble type OB-Re polypeptide can be prepared using standard bacterial and/or mammalian expression vectors, synthetically, or purified from plasma or serum,
25 all as stated in detail earlier herein. Alternatively, increased expression of native soluble OB-R polypeptide may be induce by homologous recombination techniques, as described *supra*.

Reduction of leptin activity (by developing antagonists, inhibitors, use of
30 neutralizing antibodies, or antisense molecules) should result in weight gain as might be desirable for the treatment of the weight loss associated with cancer,

AIDS or anorexia nervosa. In one embodiment, a leptin-binding form of soluble OB-R that lacks portions necessary for signal transduction or enhancement can be employed.

5 *Polypeptide-based Therapeutic Treatment*

In the simplest analysis, the *OB-R* gene is intimately associated with determination of body weight in animals, in particular, mice, rats, dogs, and man. The *OB-R* gene product, and, correspondingly, cognate molecules, appear to be part of a signaling pathway by which adipose tissue communicates with the brain and the
10 other organs. It is believed that at least one splice form of the OB-R polypeptide (*e.g.*, OB-Rb) is itself a signaling molecule, *i.e.*, a receptor for the hormone leptin.

The soluble OB-R polypeptide, or functionally active fragment thereof, or an
15 antagonist thereof, can be administered orally or parenterally, preferably parenterally. Because metabolic homeostasis is a continuous process, controlled release administration of soluble OB-R polypeptide (OB-Re) is preferred. For example, the polypeptide may be administered using intravenous infusion, an implantable osmotic pump, a transdermal patch, liposomes, or other modes of
20 administration. In one embodiment, a pump may be used [Langer *et al.*, eds., *Medical Applications of Controlled Release*, CRC Pres., Boca Raton, Florida (1974); Sefton, *CRC Crit. Ref. Biomed. Eng.*, **14**:201 (1987); Buchwald *et al.*, *Surgery*, **88**:507 (1980); Saudek *et al.*, *N. Engl. J. Med.*, **321**:574 (1989)]. In another embodiment, polymeric materials can be used [Langer, 1974, *supra*;
25 Sefton, 1987, *supra*; Smolen *et al.*, eds., *Controlled Drug Bioavailability, Drug Product Design and Performance*, Wiley, New York (1984); Ranger *et al.*, *J. Macromol. Sci. Rev. Macromol. Chem.*, **23**:61 (1983); *see also* Levy *et al.*, *Science*, **228**:190 (1985); During *et al.*, *Ann. Neurol.*, **25**:351 (1989); Howard *et al.*, *J. Neurosurg.*, **71**:105 (1989)]. In yet another embodiment, a controlled
30 release system can be placed in proximity of the therapeutic target, *i.e.*, the brain, thus requiring only a fraction of the systemic dose [*see, e.g.*, Goodson, in *Medical*

Applications of Controlled Release, vol. 2, pp. 115-138 (1984)]. Other controlled release systems are discussed in the review by Langer, *Science*, **249**:1527-1533 (1990). In another embodiment, the therapeutic compound can be delivered in a vesicle, in particular a liposome (*see* Langer, 1990 *supra*); Treat *et al.*, in

5 *Liposomes in the Therapy of Infectious Disease and Cancer*, Lopez-Berestein and Fidler (eds.), Liss, New York, pp. 353-365 (1989); Lopez-Berestein, *ibid.*, pp. 317-327; *see generally ibid.*)

In a further aspect, recombinant cells that have been transformed with the soluble splice form of the *OB-R* cDNA (*e.g.*, *OB-Re*, which will be used herein to refer to

10 a soluble *OB-R* agonist of leptin) and that express high levels of the polypeptide can be transplanted in a subject in need of enhancement of leptin activity. Preferably autologous cells transformed with *OB-Re* are transplanted to avoid rejection; alternatively, technology is available to shield non-autologous cells that

15 produce soluble factors within a polymer matrix that prevents immune recognition and rejection.

The *OB-Re* polypeptide can be delivered by intravenous, intraarterial, intraperitoneal, intramuscular, or subcutaneous routes of administration.

20 Alternatively, the *OB-Re* polypeptide, properly formulated, can be administered by nasal or oral administration. A constant supply of *OB-Re* can be ensured by providing a therapeutically effective dose (*i.e.*, a dose effective to induce metabolic changes in a subject) at the necessary intervals, *e.g.*, daily, every 12 hours, etc. These parameters will depend on the severity of the disease condition

25 being treated, other actions, such as diet modification, that are implemented, the weight, age, and sex of the subject, and other criteria, which can be readily determined according to standard good medical practice by those of skill in the art.

30 It can readily be appreciated by one of ordinary skill in the art that a soluble *OB-R* leptin antagonist can also be administered as described above for *OB-Re*.

Pharmaceutical Compositions

In yet another aspect of the present invention, provided are pharmaceutical compositions of the above. Such pharmaceutical compositions may be for administration for injection, or for oral, pulmonary, nasal or other forms of administration. In general, comprehended by the invention are pharmaceutical compositions comprising effective amounts of protein or derivative products of the invention together with pharmaceutically acceptable diluents, preservatives, solubilizers, emulsifiers, adjuvants and/or carriers. Such compositions include diluents of various buffer content (*e.g.*, Tris-HCl, acetate, phosphate), pH and ionic strength; additives such as detergents and solubilizing agents (*e.g.*, Tween 80, Polysorbate 80), anti-oxidants (*e.g.*, ascorbic acid, sodium metabisulfite), preservatives (*e.g.*, Thimersol, benzyl alcohol) and bulking substances (*e.g.*, lactose, mannitol); incorporation of the material into particulate preparations of polymeric compounds such as polylactic acid, polyglycolic acid, etc. or into liposomes. Hylauronic acid may also be used. Such compositions may influence the physical state, stability, rate of *in vivo* release, and rate of *in vivo* clearance of the present proteins and derivatives. *See, e.g.*, *Martin, Remington's Pharmaceutical Sciences*, 18th Ed. (1990, Mack Publishing Co., Easton, PA 18042) pages 1435-1712 which are herein incorporated by reference. The compositions may be prepared in liquid form, or may be in dried powder, such as lyophilized form.

Oral Delivery

Contemplated for use herein are oral solid dosage forms, which are described generally in *Martin, Remington's Pharmaceutical Sciences*, 18th Ed. (1990 Mack Publishing Co. Easton PA 18042) at Chapter 89, which is herein incorporated by reference. Solid dosage forms include tablets, capsules, pills, troches or lozenges, cachets or pellets. Also, liposomal or proteinoid encapsulation may be used to formulate the present compositions (as, for example, proteinoid microspheres reported in U.S. Patent No. 4,925,673). Liposomal encapsulation may be used and the liposomes may be derivatized with various polymers (*E.g.*, U.S. Patent

No. 5,013,556). A description of possible solid dosage forms for the therapeutic is given by Marshall, in *Modern Pharmaceutics*, Chapter 10, Banker and Rhodes ed., (1979), herein incorporated by reference. In general, the formulation will include the protein (or chemically modified protein), and inert ingredients which
5 allow for protection against the stomach environment, and release of the biologically active material in the intestine.

Also specifically contemplated are oral dosage forms of the above derivatized proteins. Protein may be chemically modified so that oral delivery of the
10 derivative is efficacious. Generally, the chemical modification contemplated is the attachment of at least one moiety to the protein (or peptide) molecule itself, where said moiety permits (a) inhibition of proteolysis; and (b) uptake into the blood stream from the stomach or intestine. Also desired is the increase in overall stability of the protein and increase in circulation time in the body. Examples of
15 such moieties include: polyethylene glycol, copolymers of ethylene glycol and propylene glycol, carboxymethyl cellulose, dextran, polyvinyl alcohol, polyvinyl pyrrolidone and polyproline. Abuchowski *et al.*, 1981, *supra*; Newmark *et al.*, *J. Appl. Biochem.*, 4:185-189 (1982). Other polymers that could be used are poly-1,3-dioxolane and poly-1,3,6-tioxocane. Preferred for pharmaceutical usage, as
20 indicated above, are polyethylene glycol moieties.

For the protein (or derivative) the location of release may be the stomach, the small intestine (the duodenum, the jejunum, or the ileum), or the large intestine. One skilled in the art has available formulations which will not dissolve in the
25 stomach, yet will release the material in the duodenum or elsewhere in the intestine. Preferably, the release will avoid the deleterious effects of the stomach environment, either by protection of the protein (or derivative) or by release of the biologically active material beyond the stomach environment, such as in the intestine.

To ensure full gastric resistance, a coating impermeable to at least pH 5.0 is essential. Examples of the more common inert ingredients that are used as enteric coatings are cellulose acetate trimellitate (CAT), hydroxypropylmethylcellulose phthalate (HPMCP), HPMCP 50, HPMCP 55, polyvinyl acetate phthalate (PVAP), Eudragit L30D, Aquateric, cellulose acetate phthalate (CAP), Eudragit L, Eudragit S, and Shellac. These coatings may be used as mixed films.

A coating or mixture of coatings can also be used on tablets, which are not intended for protection against the stomach. This can include sugar coatings, or coatings which make the tablet easier to swallow. Capsules may consist of a hard shell (such as gelatin) for delivery of dry therapeutic *i.e.* powder; for liquid forms, a soft gelatin shell may be used. The shell material of cachets could be thick starch or other edible paper. For pills, lozenges, molded tablets or tablet triturates, moist massing techniques can be used.

The therapeutic can be included in the formulation as fine multiparticulates in the form of granules or pellets of particle size about 1mm. The formulation of the material for capsule administration could also be as a powder, lightly compressed plugs or even as tablets. The therapeutic could be prepared by compression.

Colorants and flavoring agents may all be included. For example, the protein (or derivative) may be formulated (such as by liposome or microsphere encapsulation) and then further contained within an edible product, such as a refrigerated beverage containing colorants and flavoring agents.

One may dilute or increase the volume of the therapeutic with an inert material. These diluents could include carbohydrates, especially mannitol, α -lactose, anhydrous lactose, cellulose, sucrose, modified dextrans and starch. Certain inorganic salts may be also be used as fillers including calcium triphosphate, magnesium carbonate and sodium chloride. Some commercially available diluents are Fast-Flo, Emdex, STA-Rx 1500, Emcompress and Avicell.

Disintegrants may be included in the formulation of the therapeutic into a solid dosage form. Materials used as disintegrants include but are not limited to starch including the commercial disintegrant based on starch, Explotab. Sodium starch glycolate, Amberlite, sodium carboxymethylcellulose, ultramylopectin, sodium alginate, gelatin, orange peel, acid carboxymethyl cellulose, natural sponge and bentonite may all be used. Another form of the disintegrants are the insoluble cationic exchange resins. Powdered gums may be used as disintegrants and as binders and these can include powdered gums such as agar, Karaya or tragacanth.

10 Alginic acid and its sodium salt are also useful as disintegrants.

Binders may be used to hold the therapeutic agent together to form a hard tablet and include materials from natural products such as acacia, tragacanth, starch and gelatin. Others include methyl cellulose (MC), ethyl cellulose (EC) and

15 carboxymethyl cellulose (CMC). Polyvinyl pyrrolidone (PVP) and hydroxypropylmethyl cellulose (HPMC) could both be used in alcoholic solutions to granulate the therapeutic.

An antifrictional agent may be included in the formulation of the therapeutic to

20 prevent sticking during the formulation process. Lubricants may be used as a layer between the therapeutic and the die wall, and these can include but are not limited to: stearic acid including its magnesium and calcium salts, polytetrafluoroethylene (PTFE), liquid paraffin, vegetable oils and waxes. Soluble lubricants may also be used such as sodium lauryl sulfate, magnesium lauryl

25 sulfate, polyethylene glycol of various molecular weights, and Carbowax 4000 and 6000.

Glidants that might improve the flow properties of the drug during formulation and to aid rearrangement during compression might be added. The glidants may

30 include starch, talc, pyrogenic silica and hydrated silicoaluminate.

To aid dissolution of the therapeutic into the aqueous environment, a surfactant might be added as a wetting agent. Surfactants may include anionic detergents such as sodium lauryl sulfate, dioctyl sodium sulfosuccinate and dioctyl sodium sulfonate. Cationic detergents might be used and could include benzalkonium chloride or benzethonium chloride. The list of potential nonionic detergents that could be included in the formulation as surfactants are lauromacrogol 400, polyoxyl 40 stearate, polyoxyethylene hydrogenated castor oil 10, 50 and 60, glycerol monostearate, polysorbate 40, 60, 65 and 80, sucrose fatty acid ester, methyl cellulose and carboxymethyl cellulose. These surfactants could be present in the formulation of the protein or derivative either alone or as a mixture in different ratios.

Additives which potentially enhance uptake of the protein (or derivative) are for instance the fatty acids oleic acid, linoleic acid and linolenic acid.

Controlled release formulation may be desirable. The drug could be incorporated into an inert matrix which permits release by either diffusion or leaching mechanisms *i.e.*, gums. Slowly degenerating matrices may also be incorporated into the formulation. Another form of a controlled release of this therapeutic is by a method based on the Oros therapeutic system (Alza Corp.), *i.e.* the drug is enclosed in a semipermeable membrane which allows water to enter and push drug out through a single small opening due to osmotic effects. Some enteric coatings also have a delayed release effect.

Other coatings may be used for the formulation. These include a variety of sugars which could be applied in a coating pan. The therapeutic agent could also be given in a film-coated tablet; the materials used in this instance are divided into 2 groups. The first are the nonenteric materials and include methyl cellulose, ethyl cellulose, hydroxyethyl cellulose, methylhydroxy-ethyl cellulose, hydroxypropyl cellulose, hydroxypropyl-methyl cellulose, sodium carboxy-methyl cellulose,

providone and the polyethylene glycols. The second group consists of the enteric materials that are commonly esters of phthalic acid.

5 A mix of materials might be used to provide the optimum film coating. Film coating may be carried out in a pan coater or in a fluidized bed or by compression coating.

Pulmonary Delivery

Also contemplated herein is pulmonary delivery of the present soluble protein (or derivatives thereof). The protein (or derivative) is delivered to the lungs of a mammal while inhaling and traverses across the lung epithelial lining to the blood-stream. Other reports of this include Adjei *et al.*, *Pharmaceutical Research*, 7(6):565-569 (1990); Adjei *et al.*, *International Journal of Pharmaceutics*, 63:135-144 (1990) (leuprolide acetate); Braquet *et al.*, *Journal of Cardiovascular Pharmacology*, 13(suppl. 5):143-146 (1989) (endothelin-1); Hubbard *et al.*, *Annals of Internal Medicine*, 3(3):206-212 (1989) (α 1-antitrypsin); Smith *et al.*, *J. Clin. Invest.*, 84:1145-1146 (1989) (α 1-proteinase); Oswein *et al.*, "Aerosolization of Proteins", *Proceedings of Symposium on Respiratory Drug Delivery II*, Keystone, Colorado, (March 1990) (recombinant human growth hormone); Debs *et al.*, *J. Immunol.*, 140:3482-3488 (1988) and Platz *et al.*, U.S. Patent No. 5,284,656 (granulocyte colony stimulating factor). Contemplated for use in the practice of this invention are a wide range of mechanical devices designed for pulmonary delivery of therapeutic products, including but not limited to nebulizers, metered-dose inhalers, and powder inhalers, all of which are familiar to those skilled in the art.

Some specific examples of commercially available devices suitable for the practice of this invention are the Ultravent nebulizer, manufactured by Mallinckrodt, Inc., St. Louis, Missouri; the Acorn II nebulizer, manufactured by Marquest Medical Products, Englewood, Colorado; the Ventolin metered dose inhaler, manufactured

by Glaxo Inc., Research Triangle Park, North Carolina; and the Spinhaler powder inhaler, manufactured by Fisons Corp., Bedford, Massachusetts.

All such devices require the use of formulations suitable for the dispensing of
5 protein (or derivative). Typically, each formulation is specific to the type of device
employed and may involve the use of an appropriate propellant material, in
addition to the usual diluents, adjuvants and/or carriers useful in therapy. Also, the
use of liposomes, microcapsules or microspheres, inclusion complexes, or other
types of carriers is contemplated. Chemically modified protein may also be
10 prepared in different formulations depending on the type of chemical modification
or the type of device employed.

Formulations suitable for use with a nebulizer, either jet or ultrasonic, will
typically comprise protein (or derivative) dissolved in water at a concentration of
15 about 0.1 to 25 mg of biologically active protein per ml of solution. The
formulation may also include a buffer and a simple sugar (*e.g.*, for protein
stabilization and regulation of osmotic pressure). The nebulizer formulation may
also contain a surfactant, to reduce or prevent surface induced aggregation of the
protein caused by atomization of the solution in forming the aerosol.

20 Formulations for use with a metered-dose inhaler device will generally comprise a
finely divided powder containing the protein (or derivative) suspended in a
propellant with the aid of a surfactant. The propellant may be any conventional
material employed for this purpose, such as a chlorofluorocarbon, a
25 hydrochlorofluorocarbon, a hydrofluorocarbon, or a hydrocarbon, including
trichlorofluoromethane, dichlorodifluoromethane, dichlorotetrafluoroethanol, and
1,1,1,2-tetrafluoroethane, or combinations thereof. Suitable surfactants include
sorbitan trioleate and soya lecithin. Oleic acid may also be useful as a surfactant.

30 Formulations for dispensing from a powder inhaler device will comprise a finely
divided dry powder containing protein (or derivative) and may also include a

bulking agent, such as lactose, sorbitol, sucrose, or mannitol in amounts which facilitate dispersal of the powder from the device, *e.g.*, 50 to 90% by weight of the formulation. The protein (or derivative) should most advantageously be prepared in particulate form with an average particle size of less than 10 μm (or
5 microns), most preferably 0.5 to 5 μm , for most effective delivery to the distal lung.

Nasal Delivery

Nasal delivery of the protein (or derivative) is also contemplated. Nasal delivery
10 allows the passage of the protein to the blood stream directly after administering the therapeutic product to the nose, without the necessity for deposition of the product in the lung. Formulations for nasal delivery include those with dextran or cyclodextran.

15 *Methods of Treatment, Methods of Preparing a Medicament*

In yet another aspect of the present invention, methods of treatment and manufacture of a medicament are provided. Conditions alleviated by or modulated by the administration of the present derivatives are those indicated above.

20 *Dosages*

For all of the above molecules, as further studies are conducted, information will emerge regarding appropriate dosage levels for treatment of various conditions in various patients, and the ordinary skilled worker, considering the therapeutic context, age and general health of the recipient, will be able to ascertain the
25 proper dosage. Generally, for injection or infusion, dosage will be between 0.01 μg of biologically active protein/kg body weight, (calculating the mass of the protein alone, without chemical modification), and 10 mg/kg (based on the same). The dosing schedule may vary, depending on the circulation half-life of the protein or derivative used, whether the polypeptide is delivered by bolus dose or
30 continuous infusion, and the formulation used.

Administration with other compounds

For therapy associated with obesity, one may administer the present soluble protein (or derivatives) in conjunction with one or more pharmaceutical compositions used for treating other clinical complications of obesity, such as those used for treatment of diabetes (*e.g.*, insulin), high blood pressure, high cholesterol, and other adverse conditions incident to obesity. Also, other appetite suppressants may be co-administered, *e.g.*, amphetamines. Administration may be simultaneous (for example, administration of a mixture of the present protein and insulin) or may be *in seriatim*.

10

Nucleic Acid-based Therapeutic Treatment

An *OB-R* gene capable of mediating signal transduction, *e.g.*, *OB-Rb*, could be introduced into human hypothalamus cells to develop gene therapy for obesity. Such therapy would be expected to decrease body weight. Conversely, introduction of antisense constructs into brain cells, particularly hypothalamus but also including choroid plexus, or other cells where *OB-R* is expressed, would reduce the levels of active *OB-R* polypeptide and would be predicted to increase body adiposity.

20 In one embodiment, a gene encoding an *OB-R* polypeptide is introduced *in vivo* in a viral vector. Such vectors include an attenuated or defective DNA virus, such as but not limited to herpes simplex virus (HSV), papillomavirus, Epstein Barr virus (EBV), adenovirus, adeno-associated virus (AAV), and the like. Defective viruses, which entirely or almost entirely lack viral genes, are preferred:

25 Defective virus is not infective after introduction into a cell. Use of defective viral vectors allows for administration to cells in a specific, localized area, without concern that the vector can infect other cells. Thus, brain tissue can be specifically targeted. Examples of particular vectors include, but are not limited to, a defective herpes virus 1 (HSV1) vector [Kaplitt *et al.*, *Molec. Cell.*

30 *Neurosci.*, 2:320-330 (1991)], an attenuated adenovirus vector, such as the vector described by Stratford-Perricaudet *et al.*, *J. Clin. Invest.*, 90:626-630 (1992), and

a defective adeno-associated virus vector [Samulski *et al.*, *J. Virol.*, **61**:3096-3101 (1987); Samulski *et al.*, *J. Virol.*, **63**:3822-3828 (1989)].

In another embodiment, the gene can be introduced in a retroviral vector, *e.g.*, as
 5 described in Anderson *et al.*, U.S. Patent No. 5,399,346; Mann *et al.*, *Cell*,
33:153 (1983); Temin *et al.*, U.S. Patent No. 4,650,764; Temin *et al.*, U.S.
 Patent No. 4,980,289; Markowitz *et al.*, *J. Virol.*, **62**:1120 (1988); Temin *et al.*,
 U.S. Patent No. 5,124,263; International Patent Publication No. WO 95/07358,
 published March 16, 1995, by Dougherty *et al.*; and Kuo *et al.*, *Blood*, **82**:845
 10 (1993).

Alternatively, the vector can be introduced *in vivo* by lipofection. For the past
 decade, there has been increasing use of liposomes for encapsulation and
 transfection of nucleic acids *in vitro*. Synthetic cationic lipids designed to limit the
 15 difficulties and dangers encountered with liposome mediated transfection can be
 used to prepare liposomes for *in vivo* transfection of a gene encoding a marker
 [Felgner *et al.*, *Proc. Natl. Acad. Sci. USA*, **84**:7413-7417 (1987); *see* Mackey *et al.*,
Proc. Natl. Acad. Sci. USA, **85**:8027-8031 (1988)]. The use of cationic lipids
 may promote encapsulation of negatively charged nucleic acids, and also promote
 20 fusion with negatively charged cell membranes [Felgner *et al.*, *Science*,
337:387-388 (1989)]. The use of lipofection to introduce exogenous genes into
 specific organs *in vivo* has certain practical advantages. Molecular targeting of
 liposomes to specific cells represents one area of benefit. It is clear that directing
 transfection to particular cell types would be particularly advantageous in a tissue
 25 with cellular heterogeneity, such as the pancreas, liver, kidney, and brain. Lipids
 may be chemically coupled to other molecules for the purpose of targeting (*see*
 Mackey *et al.*, 1988, *supra*). Targeted peptides, *e.g.*, hormones or
 neurotransmitters, and proteins such as antibodies, or non-peptide molecules could
 be coupled to liposomes chemically.

It is also possible to introduce the vector *in vivo* as a naked DNA plasmid. Naked DNA vectors for gene therapy can be introduced into the desired host cells by methods known in the art, *e.g.*, transfection, electroporation, microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, use of a gene gun, or use of a DNA vector transporter (see, *e.g.*, Wu *et al.*, *J. Biol. Chem.*, **267**:963-967 (1992); Wu *et al.*, *J. Biol. Chem.*, **263**:14621-14624 (1988); Hartmut *et al.*, Canadian Patent Application No. 2,012,311, filed March 15, 1990).

10 Agricultural Applications

The *OB-R* gene can also be isolated from domestic animals, and the corresponding OB-R polypeptide obtained thereby. It is expected that the probe derived from the murine *OB-R* gene hybridizes to corresponding homologous coding sequences from a large number of species of animals. As discussed for human therapies, recombinant proteins can also be prepared and administered to domestic animals. Administration of the soluble polypeptide can be implemented to produce leaner food animals, such as beef cattle, swine, poultry, sheep, etc. Preferably, an autologous OB polypeptide is administered, although the invention contemplates administration of non-autologous polypeptide as well. Since the soluble OB-R polypeptide consists of approximately 800 amino acid residues, it may be highly immunogenic. Thus, administration of autologous polypeptide is preferred.

Alternatively, the introduction of the cloned genes into transgenic domestic animals would allow one to potentially decrease body weight and adiposity by overexpressing an *OB-R* transgene. The simplest means of achieving this would be to target an *OB-R* transgene to brain using its own or another brain specific promoter.

30 Conversely, increases in body fat might be desirable in other circumstances such as for the development of Kobe beef or fatty liver to make foie gras. This could

be accomplished by targeting an antisense *OB-R* transgene to brain, or by using gene knockout technology. Alternatively, where an increase in body weight at percentage of fat is desired, an inhibitor or antagonist of the OB-R polypeptide can be administered. Such inhibitors or antagonists include, but are not limited to, antibodies reactive with the polypeptide, and fragments of the polypeptide that bind but do not activate the OB receptor, *i.e.*, antagonists of leptin.

Cosmetic Implications

The OB-R polypeptide has significant value for cosmetic use, in addition to the health benefits. In particular, since the OB-R polypeptides of the invention, including derivatives and agonist analogs thereof, are useful for modulation of the rate and quantity of fat cell deposition in an animal, they are useful for reducing unsightly fat tissue, *e.g.*, fat deposits in the abdomen, hips, thighs, neck, and chin that do not necessarily amount to an obese condition, but which nevertheless detract from an individual's appearance. The fat reduction effect is thought to be accomplished, in part, by a reduction in appetite, *i.e.*, a reduction in food intake, by an increase in basal metabolism, or both. Thus, the present soluble OB-R polypeptide, or its derivatives or agonist analogs, is useful for administration to a subject to effect cosmetic changes in fat tissue deposits, whether by modulating fat deposition, reducing appetite, or both.

In addition, the present compositions and methods may be used in conjunction with various procedures, such as cosmetic surgeries designed to alter the overall appearance of a body (*e.g.*, liposuction or laser surgeries designed to reduce body mass by aspirating or ablating fat tissue), exercise (especially running and weight training), low fat diet, hypnosis, biofeedback, as examples of the ways one may attempt to decrease the percentage of fat tissue and improve the appearance of the body.

Accordingly, the present invention relates to a method for effecting cosmetic fat tissue modulation in an individual comprising administering a fat modulating amount of a soluble OB-R polypeptide, or derivative or agonist analog thereof, to an individual who desires cosmetic fat tissue modulation to improve overall body appearance. In a particular aspect, the fat tissue modulation is a consequence of appetite suppression. Preferably, the fat tissue modulation is a reduction in fat tissue.

In a further embodiment, the invention relates to a method for effecting cosmetic fat tissue loss comprising combining a procedure for changing body appearance with administration of a fat modulating amount of a soluble OB-R polypeptide, or derivative or agonist analog thereof, to an individual who desires cosmetic fat tissue modulation to improve overall body appearance.

The invention may be better understood by reference to the following Examples, which are intended to be exemplary of the invention and not limiting thereof.

EXAMPLE 1: ISOLATION OF *DB* cDNA CLONES

20

Mutations in the mouse *db* gene result in severe obesity and diabetes in a syndrome that resembles morbid human obesity [Hummel *et al.*, *Science*, **153**:1127 (1966)]. Previous data suggested that the *db* gene encoded the receptor for the gene product of the *ob* locus, known as leptin [Coleman, *Diabetologia*, **14**:141 (1978); Zhang *et al.*, *Nature*, **372**:425 (1994)]. Recently, a report that the leptin receptor was cloned from choroid plexus appeared; this clone was shown to map to the same region of chromosome 4 as *db* [Tartaglia *et al.*, *Cell*, **83**:1263 (1995)]. This receptor is a member of the family of receptors that associate with the JAK tyrosine kinases. However, mutations in this receptor were not identified in C57BL/6J *db/db* mice, suggesting that the mutation in these animals might be in a splice variant of this gene [Tartaglia *et al.*, *supra*].

The present Example shows that the leptin receptor maps to the same 300 kB interval on mouse chromosome 4 as *db*. cDNA selection and exon trapping from this region identified several ESTs with sequences identical to the leptin receptor. Characterization of the corresponding cDNA clones isolated from a mouse brain

5 cDNA library revealed that there are at least five alternatively spliced forms of the leptin receptor, each with differences at their amino and/or carboxy terminus. One of the splice variants is expressed at a high level in the hypothalamus and at a lower level in other tissues. This transcript is mutant in C57BL/Ks *db/db* mice. This mutation is the result of abnormal splicing leading to a 106 bp insertion into

10 the 3' end of the RNA, which results in a truncated cytoplasmic region that deletes "Box 2", a protein site known to interact with JAK proteins [Murakami *et al.*, *Proc. Natl. Acad. Sci. USA*, **88**:11349 (1991); Fukunaga, *et al.*, *EMBO*, **10**:2855 (1991)]; it is likely to be defective in signal transduction [Bahary *et al.*, *Proc. Natl. Acad. Sci. USA*, **87**:8642 (1990); Modl *et al.*, *Dytogenetics Cell Genetics*,

15 **67**:232 (1995)]. These data suggest that the weight reducing effects of leptin are mediated via interactions with a receptor in the hypothalamus, and perhaps other tissues.

Materials and Methods

20 *Isolation of genomic clones.* YAC clones were isolated by PCR screening and sized on a CHEF MAPPER (Bio-Rad) [Green *et al.*, *Proc. Natl. Acad. Sci. USA*, **87**:1213 (1996); Kasumi *et al.*, *Mammalian Genome*, **4**:391 (1993)]. YAC ends were recovered using vectorette PCR and plasmid end rescue [Riley *et al.*, *Nucl. Acids Res.*, **18**:2887 (1990); Hermanson *et al.*, *Nucl. Acids Res.*, **19**:4943 (1991)].

25 P1 clones were isolated by sending specific pairs of PCR primers to Genome Systems Inc. (St. Louis, MO) who provided single picks of individual mouse P1 clones [Sternsberg, *Trends Genet.*, **8**:11 (1992)]. P1 ends were recovered using vectorette PCR [Hartl *et al.*, *Bio Techniques*, **15**:201 (1993)]. BACs were isolated as described [Shizuya, *Proc. Natl. Acad. Sci. USA*, **89**:8794 (1992)].

30 Primer selection and PCR amplification were performed as described: initial denaturation

at 94° C for 3 min., 25 cycles of 94° for 1 min., 55° for 2 min. and 72° for 3 min [Zhang, 1994, *supra*.] The primers were:

D4Rck6f 5' ATCTTGGGTTCTCTGAAGAA 3' (SEQ ID NO:20);

D4Rck6r 5' GAGATTGTCAGTCACAGCCTC3' (SEQ ID NO:21);

5 D4Rck7f 5' ATCTGAATTGGAATCAAATACAC 3' (SEQ ID NO:22);

D4Rck 7r 5' AAATCTGTTATCCTTCTGAAAC 3' (SEQ ID NO:23).

Isolation of db clones. cDNA selection was performed as described using mouse brain hypothalamic RNA as the starting material [Morgan *et al.*, *Nucl. Acids Res.*,
10 20:5173 (1992). Library screening, exon trapping, and DNA sequencing were performed as described [Zhang *et al.*, 1994, *supra*, (*see Example 3*)]. The C-terminal sequences of OB-Ra, OB-Re, OB-Rd and OB-Re were found in different cDNA clones. The C-terminal sequence of OB-Rb was not full length. The C-terminus sequence of this variant was initially completed by sequencing genomic
15 DNA. The template was prepared using vectorette PCR of BAC 242 with primers from the cDNA²⁹ [Riley *et al.*, *Nucl. Acids Res.*, 18:2887 (1990)]. The sequence was confirmed by sequencing RT-PCR products.

Identification of mutations in db. RT-PCR and sequencing were performed as
20 described. Genomic sequences at the splice acceptor of OB-Rb were obtained by vectorette PCR of BAC 242 with the OB-Rb reverse primer. For RT-PCR of OB-Ra, OB-Rb, OB-Re and OB-Rd the forward primer was the same
5' ACACTGTTAATTTACACCAGAG 3' (SEQ ID NO:24) (also labeled F1 in Figure 3C). The reverse (r) primers were:
25 OB-Ra 5' AGTCATTCAAACCATTAGTTTAGG 3' (SEQ ID NO:25),
OB-Rb 5' TGGATAAACCCCTTGCTCTTCA 3' (SEQ ID NO:26),
OB-Rc 5' TGAACACAACAACATAAAGCCC 3' (SEQ ID NO:27),
OB-Rd 5' AGGCTCCCTCAGGGCCAC 3' (SEQ ID NO:28). The intron primer for OB-Rb (labeled F2 in Figure 3C) was
30 5' GTGACTGAATGAAGATGTAATATAC 3' (SEQ ID NO:29).

Tissue distribution of the alternatively spliced leptin receptor. RT-PCR was performed as described. The primer sequences for OB-Ra, OB-Rb, OB-Rc, and OB-Rd are shown above. The primers for OB-Re were:

- f 5' TGTTATATCTGGTTATTGAATGG (SEQ ID NO:30),
 5 r 5' CATTAAATGATTTATTATCAGAATTGC 3' (SEQ ID NO:31).

Results and Discussion

A series of genetic crosses segregating *db* were established. These included 50 obese (*db/db*) progeny of a C57BL/Ks *db/db* x *Mus spretus* intercross and 350
 10 obese (*db/db*) progeny of a C57BL/Ks *db/db* x *Mus castaneus* intercross. The assignment of genotype as the *db* locus was made as previously described [Bahary et al., *Proc. Nat. Acad. Sci. USA*, **37**:8642 (1990)].

Several microsatellite markers flanking *db* were used to type DNA from each
 15 animal. These included a distal marker, D4Mit31 and a proximal marker, *Ifnα*. A genetic map in the region of *db* was compiled using these and other loci (Figure 1). The mouse homologous of two previously cloned human genes were found in the region of *db*: JAK1 and PDE4B. Both of these genes map to human chromosome 1p31 suggesting that the human *db* gene maps to this chromosomal
 20 region [Modl et al., *Cytogenetics Cell Genetics*, **69**:232 (1995); Milatovich et al., *Somatic Cell Mol. Gen.*, **20**:75 (1994)].

A microdissection clone, D4Rck22, was found to be distal to *db* and recombinant in three animals [Bahary et al., *Mammalian Genome*, **4**:511-515 (1993)]. D6Rck
 25 22 was used as the starting point of a chromosome walk using yeast artificial chromosomes (YACs,) bacterial artificial chromosomes (BACs) and P1 bacteriophage clones [Zhang et al., 1994, *supra*; Steinberg, *Trends Genet.*, **8**:11 (1994); Shizuya, *Proc. Natl. Acad. Sci. USA*, **89**:8794 (1992)]. A 2.7 mB contig was assembled by chromosome walking from this marker. Of note, an
 30 approximately 200 kB region was not identified in any available mouse YAC library (~12 genome equivalents screened). This gap in the contig was closed

after chromosome walking with a series of BAC and P1 clones followed by the isolation of an additional YAC that extended an additional 500 kB proximal to this region. Recombinant animals were typed with genetic markers (both RFLPs and SSLPs) derived from the ends of the individual genomic clones. The *db* mutation was located between the distal recombination event in animals 324 and the proximal recombination events in animal 1028. Seven other proximal recombinations were noted with 50 kB, suggesting that this is a hot spot for recombination. The entire nonrecombinant interval corresponds to ~300 kB of DNA, and was spanned by two BACs, 43 and 242 (Figure 1).

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Candidate genes for *db* were isolated from BACs 43 and 242 using exon trapping and cDNA selection from mouse hypothalamus [Church *et al*, *Nature Genetics*, 6:98 (1994); Morgan *et al*, *Nucl. Acids. Res.*, 20:5173 (1992)]. A mouse brain cDNA library was screened with putative gene fragments. Analysis of eight brain cDNA clones indicated that six independent products of cDNA selection and two cDNAs identified using trapped exons were present on overlapping transcripts. The nucleotide sequence of each cDNA clone predicted N-terminal sequences at least partially identical to the mouse leptin receptor (OB-R). The position of sequences from the 5' and 3' end of the OB-R RNA was determined by the STS content of each BAC and are shown on the physical map (Figure 1). These data indicate that the gene spans ~100 kB and is transcribed toward the telomere.

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These cDNA clones diverge at the carboxy terminus. In four cases, the predicted sequences were at least partially identical up to lysine 889 of the leptin receptor, which includes the transmembrane domain. Beyond this point, the cDNAs predicts proteins with differences in the cytoplasmic domain designated OB-Ra (SEQ ID NO:11), OB-Rb (SEQ ID NO:12), OB-Rc (SEQ ID NO:13), and OB-Rd (SEQ ID NO:14) (Figure 2B). OB-Re predicted a different amino acid sequence after His⁷⁹⁶ (SEQ ID NO:15), which appears to encode a soluble receptor (Figure 2B). Clones for OB-Ra, OB-Rb, and OB-Re diverged at its N-terminus. In all

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cases, the divergent sequence also mapped to the BAC contig. OB-Ra corresponds generally to mouse OB-R [Tartaglia *et al.*, *Cell*, **83**:1263 (1995)].

The C-terminus of OB-Rb was 78 percent identical to the human OB receptor, suggesting that it is the mouse homologue [Tartaglia *et al.*, *supra*]. Leptin receptor is a member of the gp-130 family of receptors that interact with JAK protein kinase. The cytoplasmic domains of gp-130 receptors are generally required for binding JAKs and signal transduction [Kishimoto *et al.*, *Cell*, **76**:253 (1994)]. The OB-Rb cDNA sequence predicts a potential "box 2" sequence (underlined in Figure 2B), a protein motif required for binding with JAK protein kinases [Kishimoto, *supra*]. "Box 2" is conserved among many members of this receptor family and is required for signal transduction of the GCSF and IL6 receptors [Murakami *et al.*, *Proc. Natl. Acad. Sci. USA*, **88**:11349 (1991); Fukunaga *et al.*, *EMBO J.*, **10**:2855 (1991). None of the other transcripts predict a "Box 2" sequence. Of the eight cDNA clones characterized, OB-Ra was isolated three times and OB-Re two times. OB-Rb, OB-Rc, and OB-Rd were each isolated once. Additional splice variants are likely to be identified.

C57BL/Ks db/db mice have a longer fragment length of OB-Rb specific RT-PCR products (it should be noted that this PCR amplified 3' nucleic acids, and thus is specific for all splice variants having a cytoplasmic domain characteristics of OB-OB-Rb, but provides no data on the extracellular domain) from hypothalamic RNA than wild type littermates (Figure 3A). However, PCR amplified genomic DNA spanning the splice acceptor at Pro⁸⁹⁰ was of normal size in C57BL/Ks db/db compared to wild type (Figure 3B). DNA sequencing of this fragment confirmed that the genomic sequences at the splice acceptor are wild type in *db* mice. In addition, both the size and nucleotide sequence of RT-PCR products corresponding to the other 3' ends were normal in the *db* mice, suggesting that the splice donor at Lys⁸⁸⁹ is also normal (Figure 4). These data suggested that the longer OB-Rb-specific fragment from C57BL/Ks db/db mice was the result of abnormal splicing.

Sequencing of the RT-PCR products of OB-Rb from the mutant mice revealed a 106 bp insertion between the splice donor at Lys⁸⁸⁹ and splice acceptor at Pro⁸⁹⁰. The sequence of the inserted DNA was identical to the first 106 bp of the unique OB-Ra exon downstream of its splice acceptor at Arg⁸⁹⁰ (Figure 5A). Sequencing of genomic DNA and RT-PCR products from the 3' untranslated region of OB-Ra of C57BL/Ks db/db mice identified a g→t mutation 106 bp after the splice acceptor (compare Figure 5B and 5C). This mutation results in the appearance of a consensus splice donor site, AGGTAAA (Figure 5C) [Lodish *et al.*, *Mol. Cell. Biol.*, Scientific American Books : New York, pp. 1-1344 (1986)]. This mutant splice donor results in the splicing of 106 bp of the OB-Ra terminal exon into the splice acceptor at Pro⁸⁹⁰ at OB-Rb RNA. The resulting mutant OB-Rb protein has a termination codon five amino acids after the splice and an identical amino acid sequence to OB-Ra. The mutant receptor is missing most of the cytoplasmic region including the potential "Box 2" motif. While RT-PCR demonstrated that the sizes of the other 3' ends were normal in C57BL/K⁹ db/db mice, it is possible that this alternative exon is inserted into other transcripts as well.

The OB-Rb leptin receptor is expressed at a high level in the hypothalamus relative to other tissues (Figure 6). Lower level expression is seen in testes with an even lower level in adipose tissue. The other alternatively spliced mRNAs are expressed in several tissues including in some cases hypothalamus (Figure 6). OB-Re, which encodes a putative soluble receptor, is highly expressed in adipose tissue and is expressed at a lower level in brain, heart, and testes (Figure 6E).

The C57BL/Ks db/db mutation is coisogenic and results in the functional replacement of the cytoplasmic domain corresponding to OB-Rb by that of OB-Ra. These data, combined with the localization of the leptin receptor to precisely the same chromosomal region as *db*, strongly confirm that OB-Rb is allelic with *db*. The identification of mutations in the two other available alleles of *db* will provide additional information on the structure-function relationship of the protein. The fact that the C57BL/Ks db/db mutation is found in the unique C-terminus of OB-

Rb explains why the sequence of OB-Ra was unchanged in C57BL/Ks db/db mice, and that binding of leptin to the choroid plexus was normal in these animals.

Leptin binding in C57BL/Ks db/db mice is likely to be normal in all locations.

Rather, the obese phenotype appears to result from the inability of the OB-Ra C-

- 5 terminus to initiate signal transduction when expressed in place of the C-terminus of OB-Rb. Elucidation of the signal transduction pathway and identification of possible sites of JAK binding to the cytoplasmic region of this receptor are contemplated.

- 10 These results suggest that the weight reducing effects of leptin are at least partially mediated via interactions with the OB-Rb receptor having a C-terminal (cytoplasmic) domain characteristic of OB-Rb in the hypothalamus, a brain region known to play an important role in regulating body weight. This is supported by the increased potency of leptin when administered directly into the CSF and the
- 15 affects of leptin on the electrical activity of hypothalamic neurons. Leptin may modulate the activity of NPY, GLP-1 and other peptides known to affect feeding behavior in the hypothalamus and brain [Stephens *et al.*, *supra*; Tarton *et al.*, *Nature*, **379**:69 (1996)]. It may also have effects other tissues expressing the leptin receptor including fat. The receptor expressed in choroid plexus, possibly
- 20 OB-Ra or a splice variant sharing a similar C-terminus, may act to transport the protein to the CSF, a mechanism similar to that proposed for transport of insulin by the insulin receptor [Bahary *et al.*, 1990, *supra*; Partridge *et al.*, *Neurochem.*, **44**:1771 (1985); Van Houten and Posner, *Nature*, **282**:623 (1979); Wood and Park, *Am. J. Physiol.*, **233**:E331-E334 (1979)].

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OB-Re, the putative soluble receptor is believed to bind to leptin in the circulation. It could function as a transport protein to agonize leptin activity [see, *e.g.*, Davis *et al.*, *Science*, **259**:1736 (1993); Kishimoto *et al.*, *supra*; Davis *et al.*, *Science*, **260**:1805 (1993)].

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EXAMPLE 2: PREPARATION OF ANTIBODIES TO THE OB
POLYPEPTIDE

In addition to use of the recombinant protein to generate polyclonal antibodies, a
5 set of three peptide sequences from the deduced full length murine OB-R sequence
(*i.e.*, SEQ ID NOS:6-10) were identified. The four internal peptide fragments
are:

Peptide A (amino acid numbers 145-158) (SEQ ID NO:32):
10 Glu-Pro-Leu-Pro-Lys-Asn-Pro-Phe-Lys-Asn-Tyr-Asp-Ser-Lys

Peptide B (amino acid numbers 465-484) (SEQ ID NO:33):
His-Arg-Arg-Ser-Leu-Tyr-Cys-Pro-Asp-Ser-Pro-Ser-Ile-His-Pro-Thr-Ser-Glu-Pro-
Lys

15 Peptide C (amino acid numbers 863-881) (SEQ ID NO:34):
Gln-Arg-Met-Lys-Lys-Leu-Phe-Trp-Asp-Asp-Val-Pro-Asn-Pro-Lys-Asn-Cys-Ser-
Trp

20 These peptides are prepared using standard solid phase peptide synthesis. The
purified synthetic peptides are conjugated to KLH, and the peptide-KLH
conjugates are used to immunize rabbits using standard techniques. Polyclonal
antisera specific for each peptide is recovered from the rabbits.

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EXAMPLE 3: PREPARATION OF PCR PROBES FROM cDNA
SELECTION AND EXON TRAPPING CLONES

This Example describes the cDNA selection clones that were identified to
30 correspond to OB-R. PCR primers from these clones were used as probes for OB-
R cDNA and genomic clones, and are useful for identifying OB-R DNA, as well
as characterizing different OB-R splice variants.

Five cDNA selection clones were found to be useful as probes: clones 7 (SEQ ID
35 NO:35), 11 (SEQ ID NO:36), 42 (SEQ ID NO:37), 46 (SEQ ID NO:38), and 58
(SEQ ID NO:39). Two cDNA selection clones identified by hybridization with

exon trapping clones were also found to be useful probes: clones S3 (SEQ ID NO:40) and S14 (SEQ ID NO:41).

PCR primers were prepared from each of the above-noted clones for use as probes in identifying OB-R DNA. Table 1 reports the forward and reverse primers for each of the clones, and notes which splice variants of OB-R, as well as the predicted coding region, each probe labels.

Table 1. PCR Primer Probes for OB-R DNA

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Source Clone (direction)	Sequence	SEQ ID NO:	Splice variant	Recognition region
7 (forward)	CCGAGGGAATTGACAGCC	42	all	extracellular
7 (reverse)	CTCACTGTGTAGTGTGAGGAGG	43		
11 (f)	TCCTGTGGACAGAACCAGC	44	all	extracellular
11 (r)	TGACACAGCTGCTGCTCAG	45		
42 (f)	TGGATAAACCCCTTGCTCTTCA	26	b	far 3' region
42 (r)	GGTCTCAGAGCACCCAGGTA	46		
46 (f)	AGAGAGATCCCTGACCCTAGTT	47	b	extracellular
46 (r)	AACTTTCTGCCTTCTCATGTCA	48		
58 (f)	TTTCTCATCTAACAAGCAAGCA	49	b	far 5'
58 (r)	ATCTGTTTCTTGCGCAGGAT	50		
S14 (f)	CATTGTTTGGGGCTCCAG	51	d,e	extracellular
S14 (r)	AATCGTTCTGCAAATCCAGG	52		
S3 (f)	TGAAGTCATAGATGATTCGCC	53	a,d,e	extracellular
S3 (r)	GTTCGTACCCGACGTCCTG	54		

As indicated in the table, probes from clones 7 and 11 have been useful in identifying all splice forms of OB-R identified to date. Probe 42 is useful to identify a splice variant with a cytoplasmic domain corresponding to OB-Rb, *i.e.*, that is putatively signal transduction competent. Probes 46 and S14 are useful to identify splice variants having an N-terminal amino acid sequence corresponding to

OB-Rd and OB-Re (which is identical to the N-terminal sequence of the published murine OB-R up to the C-terminal splice sites identified for these proteins; *see* Figure 2B). Probe 58 is useful to identify an *OB-R* containing a unique 5' region found in the *OB-Rb* splice variant cDNA, which may be a non-coding region. S3
5 identifies nucleic acids encoding extracellular domains found in variants a, d, and e (corresponding to the published murine OB-R extracellular domain).

The hybridization conditions for screening mouse brain cDNA library were as follows: probes with a length of about 150-300bp long were labeled with
10 ³²P-dCTP using hot-PCR. The filters were first pre-hybridized for at least one hour at 65°C using RAPID-HYB buffer (Amersham LIFE SCIENCES). The labeled probe was added to a final concentration of 10⁶ cpm/ml of RAPID-HYB solution and the hybridization was done for at least 6 hours at 65°C. The filters were washed with 2X SSC/0.1%SDS, RT, for 30 min, followed by a more
15 stringent wash with 0.3X SSC/0.1%SDS, RT, for 1/2 hour.

Thus, the probes described in this example are useful for identifying OB-R, as well as identifying unique splice variants. It is believed, for example, that a splice variant with an extracellular domain corresponding to OB-Ra, or OB-Rc/d/e may
20 be joined with a cytoplasmic domain corresponding to OB-Rb.

The present invention is not to be limited in scope by the specific embodiments
25 describe herein. Indeed, various modifications of the invention in addition to those described herein will become apparent to those skilled in the art from the foregoing description and the accompanying figures. Such modifications are intended to fall within the scope of the appended claims.

30 Where nucleotide or amino acid sequence lengths are provided, or molecular weight values given, they are approximate.

Various publications are cited herein, the disclosures of which are incorporated by reference in their entireties. In particular, [Tartaglia *et al.*, *Cell*, 83:1263-1271 (1995)] is incorporated herein by reference in its entirety.